

Semester-Wise Programme structure for B.Sc. (H) Microbiology (3 Years)

Sr. No.	Year 1		Year 2		Year 3	
	Semester 1	Semester 2	Semester 3	Semester 4	Semester 5	Semester 6
1	Biochemistry-I [CU:6,L-4, P-2] {CC}	Biochemistry-II [CU:6,L-4, P-2] {CC}	Biotechniques [CU:6,L-4, P-2] {CC}	Immunology [CU:6,L-4, P-2] {CC}	Molecular Biology [CU:6,L-4, P-2] {CC}	Gene Regulation [CU:6,L-4, P-2] {CC}
2	Basic Cell Biology [CU:6,L-4, P-2] {CC}	General Bacteriology[CU:6, L-4, P-2] {CC}	Enzymology [CU:6,L-4, P-2] {CC}	Bioprocess Technology [CU:6,L-4, P-2] {CC}	Food and Dairy Technology[CU:6,L-4, P-2] {CC}	Synthetic Biology [CU:6,L-4, P-2] {CC}
3	Introduction to Microbial World [CU:4,L-3, P-1] {AC}	General Chemistry[CU:4,L-3, P-1] {AC}	Mycology and Phycology [CU:6,L-4, P-2] {CC}	Microbial Physiology and Metabolism [CU:6,L-4, P-2] {CC}	SE -I [CU:4 ,L-4] {SE} SE -II [CU:4 ,L-4] {SE}	SE -3 [CU:4 ,L-4] {SE} SE -II [CU:4 ,L-4] {SE}
4	Mathematics for Biosciences [CU:2,L-2] {SEC}	Statistics for Biosciences [CU:2,L-2] {SEC}	Protein Science [CU:4,L-4] {AC}	Recombinant DNA Technology [CU:4,L-3, P-1] {AC}	SE -2 [CU:4 ,L-4] {SE} SE -II [CU:4 ,L-4] {SE}	SE -4 [CU:4 ,L-4] {SE} SE -II [CU:4 ,L-4] {SE}
5	EVS-I [CU:2,L-2] {AEC}	EVS-II [CU:2,L-2] {AEC}	SEC-3: Programming with C [CU:2,L-1,P-1] {SEC}	SEC-4: Fundamentals of Physics [CU:2,L-2] {SEC}	SEC -5 [CU:2 ,L-2] {SEC} SEC -II [CU:2 ,L-2] {SEC}	SEC -7 [CU:2 ,L-2] {SEC} SEC -II [CU:2 ,L-2] {SEC}

6	Communication skills [CU:1,L-1] {VAC}	Communication skills [CU:1,L-1] {VAC}	-	-	SEC -6 [CU:2 ,L-2] {SEC} SEC -II [CU:2 ,L-2] {SEC}	SEC -8 [CU:2 ,L-2] {SEC} SEC -II [CU:2 ,L-2] {SEC}
7	Behavioural Sciences[CU:1, L-1] {VAC}	Behavioural Sciences[CU:1,L-1] {VAC}	-	-	-	-
8	FBL [CU:1,L-1] {VAC}	FBL [CU:1,L-1] {VAC}	-	-	-	-
9	PL/HCP [CU:1,L-1] {AEC}	PL/HCP [CU:1,L-1] {AEC}	-	-	-	-
Credits	24	24	24	24	24	24
Total Programme Credits						144

AC	Allied Course
AEC	Ability Enhancement Course
CC	Core Course
GE	General Elective
OE	Open Elective
SC	Skill component

SE	Specialization Elective Course
SEC	Skill Enhancement Course
VAC	Value Added Course
NTCC	Non Teaching Credit Course
CU	Credit Unit
L;T;P	Lecture ; Tutorial ; Practical
H	Honours

Programme structure for B.Sc. (H) Microbiology- 3 years (1st Semester)

Sr. No	Course Code	Course Title	Course Type	Credits					Credit Units
				L	T	P	FW	SW	
1	BCH101	Basic Cell Biology	Core Courses	4	0	2	0	0	6
2	BCH102	Biochemistry-I	Core Courses	4	0	2	0	0	6
3	MBO104	Introduction to Microbial World	Core Courses	3	0	1	0	0	4
4	MAT113	Mathematics for Biosciences	Skill component	2	0	0	0	0	2
5	ENV101	Environment Studies -I	Ability Enhancement Course	2	0	0	0	0	2
6	ENG101	Communication Skills	Value Added Course	1	0	0	0	0	1
7	FOL101/FOL102	Foreign Business Language	Value Added Course	1	0	0	0	0	1
8	PSY101	Behavioural Science	Value Added Course	1	0	0	0	0	1
9	INL101/INL103	Punjabi Language/Punjab History & Culture	Ability Enhancement course	1	0	0	0	0	1

Total Credits

24

BCH101: Basic Cell Biology

L	T	P	Total Credits
4	0	2	6

Course Objectives: To develop basic understanding of cell biology

Course Content and Syllabus

	Teaching Hrs
Unit I: Introduction to the Cell: theory and Broad Classification	18 hrs
Cell: The cell theory, Broad Classification of cells, Structure and function of cell organelles, Cytoskeletal structures (actin, microtubules etc.).	
Unit II: Cell wall and Cell Membrane	18 hrs
Cell wall and Cell Membrane: physical structure of model membranes in prokaryotes and eukaryotes, lipid bilayer, membrane proteins, other constituents; diffusion, osmosis, active transport, and regulation.	
Unit III: Cell division and cell cycle	18 hrs
Cell division and cell cycle: Mitosis and meiosis, Cell cycle, Apoptosis, Necrosis and Autophagy. Cell transformation and cancer: oncogenes and proto-oncogenes, Tumor suppressor genes, metastasis. Contribution of Nobel laureates in elucidation of the DNA structure, cell death and cell cycle.	
Unit IV: Cell Signalling	18 hrs
Cell signalling: General principles, signal transduction, Hormones and their receptors, second messengers, regulation of signalling pathways, bacterial chemotaxis and quorum sensing., Cell adhesion molecules, contribution in cell communication	

List of Experiments -with basic instructions

1. To study different parts of microscope
2. Cytochemical staining of proteins by Methylene blue
3. Cytochemical staining of polysaccharides by PAS
4. Study of stages of Mitosis using onion root tip
5. Study of stages of Meiosis in onion flower buds
6. Preparation of Buccal Smear for microscopic examination
7. To study the effect of isotonic, hypotonic and hypertonic solutions on cells
8. To demonstrate cell viability and cell death

Course Learning Outcomes:

- Understand types of cells and cellular organelles.
- Identify differences in the structure of different types of cell walls and membranes.
- Compare the cell division and cell cycle.
- Perceive knowledge of signalling cascades and communication networks in the cell.

Text/Reference Books

Author	Title	Publisher	Ed/year	ISBN No	Pages
De-Robertis, F.D.P. and De-Robertis Jr. E.M.F.	Cell and Molecular Biology	Lippincott Williams & Wilkins	2012	978-1469810997	---
Geoffrey, M.	The Cell: A molecular approach	Oxford Sinauer Associates, Oxford University Press	6th	978-1605351551	832
Lodish , H.F.	Molecular Cell Biology.	W H Freeman & Co	5th	978-0716788751	----

BCH102: Biochemistry-I

L	T	P	Total Credits
4	0	2	6

Course Objectives: Examine biomolecules to elucidate their structural and functional properties, fostering a deeper understanding of their roles in biological systems

Course content and syllabus

	Teaching Hours
Unit I: Water and its Properties	18 hrs
Water and its Properties: Dissociation and association constants, pH and buffers. pI, pK_a , Henderson Hasselbalch equation and its implications. Basic Thermodynamics: Laws of thermodynamics. Concepts of ΔG , ΔH and ΔS .	
Unit II: Carbohydrates	18 hrs
Carbohydrates: Structure, properties and functions of: Monosaccharides (glucose, fructose, ribose and others, D- and L- sugars, reducing and non-reducing sugars), Disaccharides (maltose, sucrose and lactose) and polysaccharides (Starch and glycogen)	
Unit III: Lipids and Nucleic Acids	18 hrs
Lipids: Classification, Structure and function. Conformation of Nucleic acids: Structural characteristics of A, B and Z-DNA. Significance of DNA and RNA.	
Unit IV: Proteins	18 hrs

<p>Proteins: Physico-chemical and structural properties of amino acids, non-protein and rare amino acids.</p> <p>Protein Structure: Primary, Secondary, Tertiary, Quaternary, structure of proteins, Forces stabilizing Primary, Secondary and Tertiary protein structures.</p> <p>Enzymes: structure & function.</p> <p>Forces that stabilize biomolecules: electrostatic and van der Waal's interaction, hydrogen bonding. Interactions with solvents, Hydrophobic effect.</p>	
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List of Practicals with basic instructions (Total = 72 hrs)

1. Preparation of solutions and buffers.
2. Preparation of 0.1M phosphate buffer, pH 7.4, 250ml without using the pH meter. (By using Henderson –Hasselbalch equation)
3. Verification of Beer Lamberts Law.
4. Estimation of carbohydrate in given solution by anthrone method.
5. Study the presence of reducing/non-reducing sugar in biological samples.
6. Protein estimation by Lowry's method and other methods.
7. Determination of acid value and saponification value of a fat.

Course Learning Outcomes:

- Understand the law of thermodynamics, water, and its properties.
- Determine the structure and properties of carbohydrates.
- Comparing the structure of various types of lipids, and their role on biological systems.
- Evaluate the structure and functional properties of proteins.

Text/Reference Books

Author	Title	Publisher	Ed/year	ISBN No	Pages
Zubay, G.L., Parson, W.W., and Vance, D.E.	Principles of Biochemistry	Wm. C. Brown, Dubuque, Iowa	1995	978-0697142757	---
Nelson, D.L. and Cox, M. Lehninger	Principles of Biochemistry	W.H.Freeman & Co Ltd	8 th /2021	978-1319381493	---

Plummer, D.T.	An Introduction to Practical Biochemistry	Tata McGraw Hill	3 rd / 2017	978- 0070994874	332
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MBO104: Introduction to Microbial World

L	T	P	Total Credits
3	0	1	4

Course Objectives: Explore the microbial world to comprehend its diversity, and foster a foundational understanding of microbiology's relevance to scientific inquiry and practical applications.

Course content and syllabus

	Teaching Hours
Unit I: Introduction to Microbiology	14 hrs
Evolution, microbial life and biosphere, Brief history of Microbiology, naming and classification of microbes, Binomial Nomenclature, Whitaker's five kingdom system, Carl Woese three kingdom of classification, Difference between prokaryotic and eukaryotic microorganisms. Introduction to bacterial taxonomy– Bergey's manual of Systematic Bacteriology (Eubacteria and Archaeobacterium).	
Unit II: The Prokaryotes: Domain Bacteria and Archaea	13 hrs
The prokaryotic groups, Domain Bacteria: Gram negative and Gram positive bacteria. Domain Archaea, diversity in archaea. Microbial Growth and Nutrition: The common nutritional requirements, Nutritional types of Microbes, Microbes and Human welfare, Microbes and Human disease.	
Unit III: The Eukaryotes- Fungi, Algae, Protozoa and Helminths	14 hrs
Algae: General characteristics of algae, Different types of life cycles in algae, Selected phyla of algae, roles of algae in nature. Lichens Fungi: General characteristics of fungi including habitat, distribution, nutritional requirements, fungal cell ultra-structure, Medically important fungi, Fungal diseases. Protozoa	

General characteristics with special reference to Amoeba, Paramecium, Plasmodium, Leishmania and Giardia, Medically important protozoa. General characteristics of slimemolds and helminths.	
Unit IV: Viruses, Viroids and Prions	13 hrs
General characteristics of Viruses, Viroids and Prions - Host range, virus size, viral structure-nucleic acid, capsid and envelope, general morphology, taxonomy of viruses, Latent and persistent viral infections, Diseases associated with viruses, viroid and prions.	

List of Experiments -with basic instructions (Total Teaching = 36 hrs)

1. Microbiology-Good Laboratory Practices and Bio-safety.
2. To study the principle and applications of important instruments (biological safety cabinets, autoclave, incubator, hot air oven, light microscope, pH meter) used in the microbiology laboratory.
3. Preparation of culture media for bacterial cultivation.
4. Sterilization of medium and glassware using Autoclave and Hot air oven, respectively and assessment for sterility.

Course Learning Outcomes:

- Understand diversity of Microbial world
- Evaluate bacterial classification and diversity
- Perceive knowledge of cellular organization of bacteria and archaea
- To understand the nutritional requirements of bacteria and different bacteriological techniques.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Pelczar, M.J. Jr., Chan ECS and Krieg, N.R.	Microbiology :Concepts and Applications	New York; Madrid: McGraw-Hill,	1993	0070492581, 97800704925 8 5	957

Cappucino, J.G.	Microbiology -A laboratory manual, 4th ed., Harlow, Addition- Wesley.	Hoboken, N.J.: Pearson	2020	0135188997, 9780135203996, 0135203996	541
Tortora GJ, Funke BR and Case CL	Microbiology: An Introduction. 9th edition	Pearson Education	2008	0805347917	912
Madigan MT, Martinko JM, Dunlap PV and Clark DP.	Brock Biology of Microorganisms	Pearson International Edition	2014	9781292018317	1030
James T. Staley Robert P. Gunsalus, Stephen Lory, Jerome J. Perry	MICROBIAL LIFE	Sinauer Associates; 2nd edition	2007	0878936858, 9780878936854	1066

MAT113: Mathematics for Biosciences

L	T	P	TOTAL CREDIT UNITS
2	0	0	2

Course Contents/syllabus:

	Teaching Hours
Unit I: Sets, Relations and Function	9 hrs
Sets and their properties, Cartesian product of Sets, relations, functions and their types and graphs	
Unit II: Matrix Algebra	9 hrs
Matrices, Types of Matrices, Addition of matrices, Subtraction of matrices and Product of matrices. Properties of Matrix Multiplication. Transpose of Matrix, Symmetric and Skew-symmetric Matrices, Inverse of Matrix and system of linear equations	
Unit III: Differential Calculus	9 hrs
Algebra of limits, Continuity, Derivative of a function, Fundamental rules for differentiation, increasing and decreasing functions, Introduction to Partial derivatives	
Unit IV: Integral Calculus	9 hrs
Indefinite and definite integrals, methods of Integration, Properties of definite integrals	

Course Learning Outcomes:

On the successful completion of this course

1. Students will demonstrate the ability to distinguish corresponding sets as representations of relations or functions by the analysis of graphical, numeric, or symbolic data
2. Students will demonstrate the ability to apply the concept of matrices in real-life situations
3. Students will understand the concepts of Limits, Continuity and Differentiability and their applications
4. Students will understand and analyze the concept of Integration with the help of Differentiation and study its various applications

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
George B. Thomas Jr., Joel Hass, Christopher Heil & Maurice D. Weir	Thomas' Calculus(14th edition)	Pearson Education	2018	978-9353060411
H.K. Dass	Higher Engineering Mathematics	S. Chand	2014	978-8121938907

ENV101: Environmental Studies -I

Course content and syllabus

L	T	P	Total Credits
2	0	0	2

	Teaching Hours
Unit-1- Multidisciplinary nature of environmental studies and Natural Resources-1	9 hrs
<i>Multidisciplinary nature of environmental studies:</i> Definition, scope and importance; components of environment –atmosphere, hydrosphere, lithosphere and biosphere. Concept of sustainability and sustainable development. <i>Natural resources:</i> Land resources and land use change, land degradation, soil erosion and desertification.	
Unit-2- Natural Resources-2	9 hrs
Deforestation: causes and impacts due to mining, dam building on environment, forests, biodiversity and tribal population. Water Resources-Use and over-exploitation of surface and groundwater, floods, drought, conflicts over water (international and inter-state). Heating of earth and circulation of air; air mass formation and precipitation. Energy resources- renewable and non-renewable energy sources, use of alternate energy sources, Growing energy needs, Case studies.	
Unit-3-Ecosystems	9 hrs

<p><i>Ecosystem</i>: What is an ecosystem; Structure and function of an ecosystem; Energy flow in the ecosystem; Food chains, food webs and ecological succession. Case studies of the following ecosystems:</p> <p>Forest ecosystem; Grassland ecosystem; Desert ecosystem; Aquatic ecosystems (ponds, streams, lakes, rivers, oceans, estuaries).</p>	
Unit-4- Biodiversity and its conservation	9 hrs
<p><i>Biodiversity</i>: Levels of biological diversity: genetic, species and ecosystem diversity; Biogeographic zones of India; biodiversity patterns and global biodiversity hot spots.</p> <p>India as a mega-biodiversity nation; endangered and endemic species of India.</p> <p>Threats to biodiversity: habitat loss, poaching of wildlife, man-wildlife conflicts, biological invasions; conservation of biodiversity: <i>in-situ</i> and <i>ex-situ</i> conservation of biodiversity.</p> <p>Ecosystem and biodiversity services: ecological, economic, social, ethical, aesthetic and information value.</p>	

Course Learning Outcomes:

1. Understand natural resources and evaluate limitations surrounding renewable and non-renewable resources
2. Understand the nuances of ecosystem and learn about behaviour of various ecosystem
3. Learn about the types, services and threats to our biodiversity and importance of conserving it.

Text/Reference Books

AUTHOR	TITLE	Publisher	Year of publication	ISBN	Pages
William P. Cunningham, MaryAnn Cunningham	Principles of Environmental Science	McGraw-Hill	2019	9781260219715	--
Dash and Dash	Fundamentals of ecology	Tata McGraw-Hill Education	2009	978-0070083660	--

<p>William P. Cunningham, Mary Ann Cunningham, Barbara Woodworth Saigo</p>	<p>Environmental Science: A global concern,</p>	<p>McGraw-Hill</p>	<p>2021</p>	<p>9781260363821</p>	<p>--</p>
<p>Gaston K.J. and Spicer, J. I.</p>	<p>Biodiversity – An Introduction 2nd edition</p>	<p>Blackwell Publishing</p>	<p>2004</p>	<p>978-1-405-11857-6</p>	<p>--</p>

ENG101: Communication Skills-I

L	T	P	Total Credits
1	0	0	1

Course Contents/syllabus:

	Teaching hours
Unit I: Basic Concepts in Communication	4 hrs
Definition of communication, Nature and process of communication, role and purpose of communication, types and channels of communication, communication networks/flow of communication: vertical, diagonal, horizontal, barriers to communication: physical, language, and semantic, socio-psychological, organizational, gateway to effective communication, towards communicative competence, choosing the appropriate channel and medium of communication, social communication: small talk and building rapport, barriers in communication.	
Unit II: Communication Types	5 hrs
Verbal communication: Oral Communication: Forms, Advantages & Disadvantages, Written Communication: Forms, Advantages & Disadvantages, Introduction of Communication Skills (Listening, Speaking, Reading, Writing), Nonverbal communication: functions and effective use, KOPPACT(Kinesics, Oculesics, Proxemics,	
Para-language, Artifacts, Chronemics, Tactilics). The implication of appropriate communication; effective ways of using social media, importance of digital literacy.	
Unit III: Reading and Writing Skills	4 hrs

Significance of reading; Reading Comprehension, gathering ideas from a given text, identify the main purpose and context of the text, evaluating the ideas, interpretation of the text, Paragraph development; essay writing.	
Unit IV: Speaking and Presentation Skills	5 hrs
Speaking skills: fluency, vocabulary, grammar, and pronunciation; effective speaking: selection of words, your voice, and non-verbal communication, functions of speaking: interaction, transaction, and performance; structuring the message; effective speaking strategies. Planning, preparation, practice, and performance; audience analysis, audio- visual aids, analyzing the non-verbal communication, methods of delivery: impromptu, extemporaneous, memorization, manuscript, and outlining.	

Course Learning Outcomes:

1. Students will be able to understand the basic processes of communication, both verbal as well as non-verbal—nature, scope, and power of communication processes.
2. Students will be able to demonstrate cultural sensitivity in communication and appreciation of cultural variations of diverse socio-cultural contexts.
3. Students will be able to develop an awareness of the role of mass media in shaping public psyche, beliefs, and perceptions about social realities and build an informed and critical perspective.
4. Students will be able to analyze situations and audiences to make right choices about the most effective and efficient ways to communicate and deliver messages.
5. Students will be able to assess various barriers in communication and develop communicative competence thereby for effective communication.

Books/literature

AUTHOR	TITLE	Publisher	Year of publication	ISBN
P. D. Chaturvedi and Mukesh Chaturvedi	Business Communication : Concepts, Cases and Applications	Pearson Education	2006	9788131701720
Meenakshi Raman and Prakash Singh	Business Communication	Oxford University Press	2012	9780198077053

Jeff Butterfield	Soft Skills for Everyone	Cengage Learning	2017	9789353 501051
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FOL101: Introduction to French Culture & Language

L	T	P	Total Credits
1	0	0	1

Course Contents/syllabus:

	Teaching hours
Unit-I Introduction to French language	4 hrs
<ul style="list-style-type: none">• Brief introduction of French and Francophone countries• Presenting oneself• Getting information about someone else• Greeting and taking leave• Asking/giving personal information	
Unit-II- A rendez-vous ; Visiting a place	5 hrs
<ul style="list-style-type: none">• Pronouncing and writing numbers in French• Spell and count numbers• Telling the time• Temporal expressions• Communicating in class• Fixing an hour, place for a meeting.• Describing a person.• Identifying a person, object and place	

<ul style="list-style-type: none"> • Describing relation in a family • A specific person, object and place 	
Unit-III- An interview	4 hrs
<ul style="list-style-type: none"> • Description of objects, people and places • Nationalities • Speaking about one's professions • Expressing Actions using regular –er ending verbs; avoir, être; reflexive verbs –usage, conjugation • Interview of celebrity 	
Unit-IV- At the discotheque	5 hrs
<ul style="list-style-type: none"> • Portrait by a journalist • Giving a positive or negative reply • Asking questions • Discussion with a person • Activities in a day 	

Course Learning Outcomes: At the end of this course, the students will be able to express themselves in writing and orally in basic French. This course content focuses on the speech of the students in a lucid and a concurrent manner using appropriate vocabulary and pronunciation techniques. Extra stress will be given on their understanding of grammatical structures and the foreign accent of the language. At the end of the course, the student shall be able to :

1. Understand information; Express in his own words; Paraphrase; Interpret and translate.
2. Apply information in a new way in a practical context
3. Analyse and break-down information to create new ideas
4. Evaluate and express opinion in a given context

Text / Reference Books:

Author	Title	Publisher	Year	ISBN No
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Christine Andant, Chaterine Metton, Annabelle Nachon, Fabienne Nugue	A Propos - A1 Livre De L'Eleve, Cahier D' Exercices	Langers International Private Limited	2010	978-9380809069
Manjiri Khandekar and Roopa Luktuke	Jumelage - 1 Methode De Fraincais - French	Langers International Private Limited	2020	978- 938080 9854
Michael Magne, Marie-Laure Lions-Olivieri	Version Originale 1: Cahier d'exercices	Maison Des Langues	2010	978848 443561 7

FOL102: Introduction to German Culture & Language

L	T	P	Total Credits
1	0	0	1

Course Contents/syllabus:

	Teaching hours
Unit-I Introduction to German Language (Einführung)	3 hrs
Introduction to German as a global language, Self-introduction and Greetings, Die Alphabeten, Phonetics: the sound of consonants and vowels, Wie buchstabieren Sie Ihren Name?	
Unit-II- Numbers and everyday conversation (die Zahl und Gespräche)	6 hrs
Counting in German from 1-100, Simple Calculation and verb 'kosten' - Wie viel kostet das? Plural Forms, Vocabulary: Wochentage, Monate, Jahreszeiten, Ordinal numbers and the question - Wann haben Sie Geburtstag?	
Unit-III- Regular verbs and nominative case: articles and pronouns (Regelmässige Verben und Nominativ Kasus: Artikel und Pronomen)	4.5 hrs
Introduction to all personal pronouns and conjugation of Regular verbs Detailed exercise on regular verbs. Reading a text on regular verbs. Introduction to definite. Vocabulary: Schulsachen und Getränke, Nominative case/ Articles (der, die, das) Nominative Pronouns: - Applicability of pronouns for both persons and things. Usage of nominative Personal Pronouns Introduction of nominative possessive pronouns usage of nominative possessive pronouns	
Unit-IV- The Family, Work-life and Professions (Familienmitglieder und Berufe) & Interrogative sentences (W-Fragen)	4.5 hrs

<p>The Family, Work-life and Professions (Familienmitglieder und Berufe)</p> <p>Vocabulary: Professions and conjugation of the verb 'sein' Introduction to simple possessive pronouns with the help of the verb 'haben' Usage of possessive pronouns. Interrogative sentences (W-Fragen) W-Fragen: who, what, where, when, which, how, how many, how much, etc. Exercises on the question pronouns</p>	
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Course Learning Outcomes: At the end of this course, the students will be able to express themselves in writing and orally in basic German. This course content focuses on the speech of the students in a lucid and a concurrent manner using appropriate vocabulary and pronunciation techniques. Extra stress will be given on their understanding of grammatical structures and the foreign accent of the language. At the end of the course, the student shall be able to:

1. Understand information; Express in his own words; Paraphrase; Interpret and translate.
2. Apply information in a new way in a practical context
3. Analyse and break-down information to create new ideas
4. Evaluate and express opinion in a given context

Text / Reference Books:

Author	Title	Publisher	Year	ISBN
Rolf Bruseke	Starten Wir A 1	Langers International Pvt Ltd (Max Hueber Verlag)	2017	978-3190160006
Giorgio Motta	Wir Plus Grundkurs Deutsch fur Junge Lerner Book	Ernst Kleit Verlag	2011	978-8183072120
Heimy Taylor, Werner Haas	Station en Deutsch Self StudyCourse German Guide	Wiley	2007	978-0470165515

PSY101: Behavioural Science: Understanding Self for Effectiveness

L	T	P	Total Credits
1	0	0	1

Course Contents/syllabus:

	Teaching Hrs
Unit I: Self: Core Competency	4.5 hrs
Understanding of Self, Components of Self – Self identity , Self concept, Self confidence, Self image , BIG5 Factors	
Unit II: Techniques of Self Awareness	4.5 hrs
Exploration through Johari Window, Mapping the key characteristics of self, Framing a charter for self Stages – self awareness, self acceptance and self realization	
Unit III: Self Esteem & Effectiveness	4.5 hrs
Meaning, Importance, Components of self esteem, High and low self esteem, Measuring your self esteem	
Unit IV: Building Positive Attitude and Emotional Competence	4.5 hrs
Meaning and nature of attitude, Components and Types of attitude, Importance and relevance of attitude Emotional Intelligence – Meaning, components, Importance and Relevance Positive and negative emotions, Healthy and Unhealthy expression of emotions	

Course Learning Outcomes: At the end of this course, the students will be able to:

1. The student will apply self-introspection as a tool for self-awareness.
2. The student will understand self-concept for self-recognition, self-improvement and perception of others.
3. The student will be able to analyze their physical self, social self, the competent self and psychological self.
4. The student will be able to analyze what motivates his/her actions and the actions of others

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
Singh A.	Achieving Behavioural Excellence for Success	Wiley Publication	2012	978812658027
Towers, Marc	Self Esteem	American Media	1995	9781884926297
Pedler Mike, Burgoyne John, Boydell Tom	A Manager's Guide to Self-Development	McGraw-Hill	2006	978-0077114701
Covey, R. Stephen	Seven habits of Highly Effective People	Simon & Schuster Ltd	2013	978-1451639612
Khera Shiv	You Can Win	Macmillan	2005	978-0333937402
Gegax Tom	Winning in the Game of Life	Harmony Books	1999	978-0609603925

Singh, Dalip	Emotional Intelligence at Work	Publications	2006	9780761935322
Goleman, Daniel	Emotional Intelligence	Bantam Books	2007	9780553095036
Goleman, Daniel	Living with E.I	Bantam Books	1998	9780553104622

INL101: Punjabi Language -I

L	T	P	Total Credits
1	0	0	1

Course content and syllabus

	Weightage (%)	Teaching Hours
Unit I:	25%	4 hours
ਆਧੁਨਿਕ ਪੰਜਾਬੀ ਕਵਿਤਾ ਦਾ ਅਧਿਐਨ (ਕਾਵਿ-ਸੁਮੇਲ ਪਾਠ-ਪੁਸਤਕ) ਕਵਿਤਾ ਦਾ ਸਾਰ/ਕੇਂਦਰੀ ਭਾਵ ਅਤੇ ਪ੍ਰਸੰਗ ਸਾਹਿਤ ਵਿਆਖਿਆ ਕਵੀ ਦੇ ਜੀਵਨ ਅਤੇ ਸਾਹਿਤਕ ਯੋਗਦਾਨ ਬਾਰੇ ਮੁੱਢਲੀ ਜਾਣਕਾਰੀ		
Unit II:	25%	4 hours
1.ਲੇਖ-ਰਚਨਾ ਲੇਖ-ਰਚਨਾ: ਮਹੱਤਵ, ਕਿਸਮਾਂ ਅਤੇ ਵੱਖ-ਵੱਖ ਵਿਸ਼ਿਆਂ ਅਨੁਸਾਰ ਵਿਹਾਰਕ ਅਭਿਆਸ 2.ਸੰਖੇਪ-ਰਚਨਾ ਸੰਖੇਪ-ਰਚਨਾ: ਮਹੱਤਵ ਅਤੇ ਤਕਨੀਕ		
Unit III:	25%	5 hours
ਵਿਆਕਰਨ ਸਿਧਾਂਤ ਅਤੇ ਵਿਹਾਰ: 1.ਵਿਆਕਰਨ: ਪਰਿਭਾਸ਼ਾ;ਮਹੱਤਤਾ;ਉਦੇਸ਼;ਵਿਆਕਰਨ ਦੇ ਅੰਗ 2. ਪੰਜਾਬੀ ਯੁਨੀਵਿਓਂਟ: ਸ੍ਰੀ ਅਤੇ ਵਿਅੰਜਨ ਯੁਨੀਆਂ ਦਾ ਵਰਗੀਕਰਨ, ਉਚਾਰਨ ਅੰਗ		
Unit IV:	25%	5 hours
ਵਿਆਕਰਨ ਸਿਧਾਂਤ ਅਤੇ ਵਿਹਾਰ: ਸ਼ਬਦ ਸ਼੍ਰੇਣੀਆਂ: ਪਰਿਭਾਸ਼ਾ ਅਤੇ ਪ੍ਰਕਾਰ ਨਾਂਵ, ਪੜਨਾਂਵ, ਵਿਸ਼ੇਸ਼ਣ, ਕਿਰਿਆ, ਕਿਰਿਆ ਵਿਸ਼ੇਸ਼ਣ, ਸਬੰਧਕ,ਯੋਜਕ ਅਤੇ ਪ੍ਰਸ਼ਨ-ਸੂਚਕ ਸ਼ਬਦ		

Course Learning Outcomes:

1. Understand modern Punjabi Poetry.
2. Interpret the importance of essay and precise writing
3. Analyze the Punjabi language structure and grammar.
4. Examine the impact and importance of grammar and language structure.

Pedagogy for Course Delivery

Lectures: 14 sessions

Presentation / Seminar/ Assignment: 2 sessions

Mid Term Test & End Term Exam: 2 sessions

Quiz: 3

Total:18 sessions

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN	Pages
ਡਾ. ਕਰਮਜੀਤ ਸਿੰਘ (ਸੰਪਾ.),	ਕਾਵਿ ਸੁਮੇਲ	ਪਬਲੀਕੇਸ਼ਨ ਬਿਊਰੋ, ਪੰਜਾਬ ਯੂਨੀਵਰਸਿਟੀ ਚੰਡੀਗੜ੍ਹ	2020	-	-
ਸੁਰਿੰਦਰ ਸਿੰਘ ਖਹਿਰਾ (ਸੰਪਾ.),	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਵਿਆਕਰਨ ਅਤੇ ਬਣਤਰ	ਪਬਲੀਕੇਸ਼ਨ ਬਿਊਰੋ, ਪੰਜਾਬੀ ਯੂਨੀਵਰਸਿਟੀ ਪਟਿਆਲਾ	2015	-	-
ਡਾ. ਹਰਕੀਰਤ ਸਿੰਘ,	ਕਾਲਜ ਪੰਜਾਬੀ ਵਿਆਕਰਨ ਅਤੇ ਲੇਖ ਰਚਨਾ	ਪੰਜਾਬ ਸਟੇਟ ਯੂਨੀਵਰਸਿਟੀ ਟੈਕਸਟ ਬੁੱਕ ਬੋਰਡ, ਚੰਡੀਗੜ੍ਹ	1999	-	-
ਡਾ. ਪ੍ਰੇਮ ਪ੍ਰਕਾਸ਼ ਸਿੰਘ	ਕਾਲਜ ਪੰਜਾਬੀ ਵਿਆਕਰਨ ਅਤੇ ਲੇਖ ਰਚਨਾ	ਮਦਾਨ ਪਬਲੀਕੇਸ਼ਨਜ਼, ਪਟਿਆਲਾ	2002	-	-
ਡਾ. ਬੂਟਾ ਸਿੰਘ ਬਰਾੜ	ਪੰਜਾਬੀ ਵਿਆਕਰਨ	ਚੇਤਨਾ ਪ੍ਰਕਾਸ਼ਨ, ਪੰਜਾਬੀ ਭਵਨ, ਲੁਧਿਆਣਾ	2012	-	-

	ਸਿਧਾਂਤ ਅਤੇ ਵਿਹਾਰ				
ਡਾ. ਬੂਟਾ ਸਿੰਘ ਬਰਾੜ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਸ਼੍ਰੇਣੀ ਅਤੇ ਸਰੂਪ	, ਵਾਰਿਸ ਸ਼ਾਹ ਫਾਊਂਡੇਸ਼ਨ, ਅੰਮ੍ਰਿਤਸਰ	2012	-	-
ਦੁਨੀ ਚੰਦ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਦਾ ਵਿਆਕਰਣ	, ਪੰਜਾਬ ਯੂਨੀਵਰਸਿਟੀ ਪਬਲੀਕੇਸ਼ਨ ਬਿਊਰੋ, ਚੰਡੀਗੜ੍ਹ	1995	-	-
ਜੋਗਿੰਦਰ ਸਿੰਘ ਪੁਆਰ ਅਤੇ ਹੋਰ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਦਾ ਵਿਆਕਰਣ (ਭਾਗ 1,2,3),	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਅਕਾਦਮੀ ਜਲੰਧਰ	2003	-	-
ਸੁਖਵਿੰਦਰ ਸਿੰਘ ਸੰਘਾ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਵਿਗਿਆਨ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਅਕਾਦਮੀ ਜਲੰਧਰ	2010	--	-
ਅਗਨੀਹੋਤਰੀ, ਵੇਦ	ਪਰਿਚਾਇਕ ਭਾਸ਼ਾ ਵਿਗਿਆਨ	ਦੀਪਕ ਪਬਲਿਸ਼ਰਜ਼ ਜਲੰਧਰ	1981		

INL102: History and Culture of Punjab

L	T	P	Total Credits
1	0	0	1

Course Contents/syllabus

	Teaching hours
Unit I:	4.5 hrs
1. Harappan Civilization: extent and town planning and socio-economic life. 2. Life in Vedic Age: socio-economic and religious; 3. Growth and impact of Jainism and Buddhism in Panjab.	
Unit II:	4.5 hrs
1. Society and Culture under Maurayas and Guptas. 2. Bhakti movement: Main features; prominent saints and their contribution. 3. Origin and development of Sufism	
Unit III:	4.5 hrs
1. Evolution of Sikhism: teaching of Guru Nanak; Institutional Development- Manji, Masand, Sangat and Pangat 2. Transformation of Sikhism: Martyrdom of Guru Arjan; New policy of Guru Hargobind, martyrdom of Guru Tegh Bahadur. 3. Institution of Khalsa: New baptism; significance	
Unit IV:	4.5 hrs

<p>1. Changes in Society in 18th century: social unrest; emergence of misls and other institutions - rakhi, gurmata, dal khalsa.</p> <p>2. Society and Culture under Maharaja Ranjit Singh.</p> <p>3. MAP (of undivided physical geographical map of Punjab): Major Historical Places: Harappa, Mohenjodaro, Sanghol, Ropar, Lahore, Amritsar, Kiratpur, Anandpur Sahib,</p> <p>Tarn Taran, Machhiwara, Goindwal, Khadur Sahib.</p>	
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Course Learning Outcomes:

1. Understand the history of various cultures in Punjab.
2. Interpret the importance of Maurayan, Gupta, and Bhakti influences on Punjab
3. Apply the teaching of Sikhism on the emergence of the Khalsa .
4. Examine the impact societal changes on socio-cultural and physical landscape of Punjab

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No
L.M Joshi,	History and Culture of the Punjab, Part-I	Punjabi University, Patiala	1989, 3 rd	-
Buddha Prakash	Glimpses of Ancient Punjab	Punjabi University, Patiala,	1983	-
Khushwant Singh	A History of the Sikhs, vol I: 1469-1839,	oxford University Press, Delhi	1991	-

Programme structure B.Sc. (H) Microbiology- 3 years (2nd Semester)

Sr. No	Course Code	Course Title	Course Type	Credits					Credit Units
				L	T	P	FW	SW	
1	BCH104	Biochemistry – II	Core Course	4	0	2	0	0	6
2	MBO103	General Bacteriology	Core Course	4	0	2	0	0	6
3	CHE113	General Chemistry	Allied Course	3	0	1	0	0	4
4	MAT114	Statistics for Biosciences	Skill Enhancement course	2	0	0	0	0	2
5	ENV106	Environmental Studies-II	Ability Enhancement Course	2	0	0	0	0	2
6	ENG103	Communication Skills -II	Value Added Course	1	0	0	0	0	1
7	PSY106	Behavioural Science - II	Value Added Course	1	0	0	0	0	1
8	FOL103/ FOL104	Foreign Business Language –II	Value Added Course	1	0	0	0	0	1
9	INL104/ INL106	Punjabi Language/ History and Culture of Punjab	Ability Enhancement Course	1	0	0	0	0	1

Total Credits

24

BCH104: Biochemistry-II

L	T	P	Total Credits
4	0	2	6

Course Objective: Delve into the biochemistry of metabolism to analyze metabolic pathways, understand energy transformation processes, and explore the regulation of biochemical reactions, fostering insights into cellular physiology and disease mechanisms.

Course content and syllabus

	Teaching Hours
Unit I: Carbohydrate Metabolism	18 hrs
Glycolytic pathway - aerobic and anaerobic glycolysis, Gluconeogenesis, Regulation of glycogen metabolism, Citric acid cycle and its regulation, Pentose phosphate pathway, Glyoxylate cycle, fate of absorbed carbohydrates, fructose, galactose, and mannose.	
Unit II: Lipid metabolism	18 hrs
Oxidation of fatty acids - Beta oxidation, alpha-oxidation, and omega oxidation, Ketogenesis, Biosynthesis of saturated and unsaturated fatty acids, Biosynthesis and degradation of tri-acyl glycerol and cholesterol, fate of absorbed dietary lipids	
Module III Protein Metabolism	18 hrs
Catabolism of amino acids, trans-amination, Oxidative and non-oxidative de-amination, Decarboxylation- urea cycle and its regulation, Biosynthesis of creatinine, fate of dietary proteins	
Module IV Nucleic Acid Metabolism & Integration of metabolic pathways	18 hrs
Catabolism and biosynthesis of nucleotides, de-novo synthesis and salvage pathways, Regulation of purine and pyrimidine biosynthesis, Interrelationship among carbohydrate, protein and fat metabolism	

List of Experiments -with basic instructions (Total Teaching = 72 hrs)

1. Ninhydrin Test for Qualitative identification of Amino acids
2. Xanthoproteic Test for Qualitative identification of Aromatic Amino acids
3. Saponification test for lipid

4. Determination of Iodine number of fatty acids
5. Estimation of cholesterol
6. Estimation of protein by Bradford/Lowry's method
7. Estimation of DNA by Di-phenyl amine (DPA) method
8. Estimation of RNA by Orcinol method

Course Learning Outcomes:

1. Students will understand the metabolic pathways linked with a series of chemical reactions occurring within a cell.
2. This course will describe the chemical changes catalyzed by cellular components and various intracellular controls.
3. Have knowledge of cellular metabolism, including central catabolic and anabolic pathways
4. Understand how different control mechanisms may be integrated to coordinate cell metabolism and function.
5. Understand how metabolism is coordinated in body systems and have knowledge of how disturbances in metabolism contribute to diseases

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
David L Nelson; and Michael M. Cox, W.H. Freeman	Lehninger's Principles of Biochemistry	WH Freeman	2012	0070492581, 9780070492585	957
Jeremy M. Berg, Lubert Stryer, John L Tymoczko, and Gregory J. Gatto,	Biochemistry	W.H. Freeman Company	2015	1319114652	1208

MBO103: General Bacteriology

L	T	P	Total Credits
4	0	2	6

Course Objectives: Study general bacteriology to comprehend bacterial morphology, physiology, and ecology, enabling the understanding of microbial roles in health, disease, and ecosystem dynamics, and the application of bacteriological knowledge in various scientific and industrial contexts.

Course content and syllabus

	Teaching Hours
Unit I: Anatomy of Prokaryotic cell	18 hrs
Cell size, shape and arrangement, glycocalyx, capsule, flagella, endoflagella, fimbriae and pili. Cell-wall: Composition and detailed structure of Gram-positive and Gram-negative cell walls, Archaeobacterial cell wall, Gram and acid fast staining mechanisms, lipopolysaccharide(LPS), sphaeroplasts, protoplasts, and L-forms. Effect of antibiotics and enzymes on the cell wall. Cell Membrane: Structure, function and chemical composition of bacterial and archaeal cell membranes.	
Cytoplasm: Ribosomes, mesosomes, inclusion bodies, nucleoid, chromosome and plasmids Endospore: Structure, formation, stages of sporulation.	
Unit II: Bacterial Taxonomy	18 hrs
Aim and principles of classification, systematics and taxonomy, concept of species, taxa, strain; conventional, molecular and recent approaches to polyphasic bacterial taxonomy, evolutionary chronometers, rRNA oligonucleotide sequencing, signature sequences, and protein sequences. Differences between eubacteria and archaeobacteria, Important archaeal and eubacterial groups.	

Unit III: Sterilization, disinfection and microscopy	18 hrs
Sterilisation and disinfection- Definitions, Principles. Methods of sterilization- Physical methods (Heat, Filtration), Radiation and Chemical methods. Control of sterilization and Testing of sterility. Microscopy – Principles, Light microscope, Phase Contrast, Dark field, Bright field, Fluorescent, Interference microscope (Stereo microscope), Confocal, Inverted microscope, and Electron microscope (TEM and SEM). Measurement of Microorganisms- Micrometry. Staining- Simple, Gram staining, Negative staining, Capsule staining, Spore staining, Flagellar staining, Nuclear staining and Acid fast staining.	
Unit IV: Bacterial nutrition and bacteriological techniques	18 hrs
Culture media: components of media, natural and synthetic media, chemically defined media, complex media, selective, differential, indicator, enriched and enrichment media. Pure culture isolation: Streaking, serial dilution and plating Selective and differential media. Growth curve and growth kinetics. Influence of environmental factors for microbial growth. Nutritional groups of bacteria: overview. Estimation of Microbes- Direct Microscopic count, Turbidometric assay, TVC- Indirect Method- CO ₂ liberation- Protein estimation- Maintenance and Preservation of cultures.	

List of Experiments -with basic instructions (Total Teaching = 72 hrs)

1. Preparation of different media: Complex media-Nutrient agar, McConkey agar, EMB agar.
2. Isolation and purification of micro-organisms from soil/water/air.
3. Estimation of CFU count by spread plate method/pour plate method.
4. Isolation of pure cultures of bacteria by streaking method.
5. Preservation of bacterial cultures by various techniques.
6. To perform the Simple staining and negative staining of a pure bacterial culture
7. To perform the Gram's staining and acid fast staining of a pure bacterial culture

Course Learning Outcomes:

1. Understand diversity of Microbial world
2. Evaluate bacterial classification and diversity
3. Perceive knowledge of cellular organization of bacteria and archaea
4. To understand the nutritional requirements of bacteria and different bacteriological techniques.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
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Pelczar, M.J.Jr., Chan ECS and Krieg, N.R.	Microbiology :Concepts and Applications	New York; Madrid: McGraw-Hill,	1993	0070492581, 9780070492585	957
Cappucino, J.G.	Microbiology -A laboratory manual, 4th ed., Harlow, Addition-Wesley.	Hoboken, N.J.: Pearson	2020	0135188997, 9780135203996,	541
Tortora GJ, Funke BR and Case CL	Microbiology :An Introduction. 9th edition.	Pearson Education	2008	0805347917	912
Madigan MT, Martinko JM, Dunlap PV and Clark DP.	Brock Biology of Microorganisms	Pearson International Edition	2014	9781292018317	1030
James T. Staley Robert P. Gunsalus Stephen lory, Jerome J. Perry	Microbial Life	Sinauer Associates; 2nd edition	2007	0878936858, 9780878936854	1066

CHE112: General Chemistry

L	T	P	SW/ FW	TOTAL CREDIT UNITS
3	0	1	0	4

Course Objectives: To develop basic understanding of atomic structure and related physico-chemical properties of elements of the periodic table. In addition, this course focuses on fundamentals of organic molecules, structure, bonding, reactivity and reaction mechanisms as well as familiarization with various states of matter and its physical laws related to describe them.

Course Content

	Teaching hours
Unit I: The Atomic Theory	14 h
Bohr's theory, Wave mechanics: de' Broglie equation, Heisenberg's Uncertainty Principle and its significance, Schrödinger's wave equation, Quantum numbers and their significance Radial and angular wave functions for hydrogen atom. Radial and angular distribution curves. Shapes of s, p, d and f orbitals. Pauli's Exclusion Principle, Hund's rule of maximum multiplicity, Aufbau's principle and its limitations, Variation of orbital energy with atomic number.	
Unit II: The Periodic Table: History and Periodic Trends	14 h
Detailed discussion of the following properties of s, p, d, f block elements in long form of periodic table. with reference to s and p-block; Effective nuclear charge, shielding or screening effect, Slater rules, variation of effective nuclear charge in periodic table, Atomic radii (van'der Waals), Ionic and crystal radii (octahedral and tetrahedral) Covalent radius, Ionization enthalpy and factors affecting successive ionization energies. Application of ionization enthalpy, trends in electron gain enthalpy electronegativity- Pauling, Mullika Allred Rochow scales, electro-negativity and bond order, partial charge, hybridization, group electronegativity.	
Unit III: Fundamentals of Organic Chemistry	13 h

Classification, and Nomenclature, Hybridization, Shapes of molecules, Influence of hybridization on bond properties. Electronic Displacements: Inductive, electromeric, resonance and mesomeric effects, hyperconjugation and their applications; Dipole moment; Organic acids and bases; their relative strength. Homolytic and Heterolytic fission with suitable examples. Curly arrow rules, formal charges; Electrophiles and Nucleophiles; Nucleophilicity and basicity; Types, shape and relative stabilities of reaction intermediates (Carbocations, Carbanions, Free radicals and Carbenes). Organic reactions and their mechanism: Addition, Elimination and Substitution reactions.	
Unit IV: Gaseous state of matter	13 h
Deviations from ideal gas behavior, compressibility factor, and its variation with pressure for different gases. Causes of deviation from ideal behavior. van der Waals equation of state, its derivation and application in explaining real gas behaviour; van der Waals equation expressed in virial form, Boyle temperature. Isotherms of real gases and their comparison with van der Waals isotherms, continuity of states, critical state, critical and van der Waals constants, law of corresponding states. Kinetic molecular model of a gas: postulates and derivation of the kinetic gas equation; collision frequency; collision diameter; mean free path and viscosity of gases, including their temperature and pressure dependence, relation between mean free path and coefficient of viscosity, calculation of σ from η ; variation of viscosity with temperature and pressure. Maxwell distribution and its use in evaluating molecular velocities (average, root mean square and most probable) and average kinetic energy, law of equipartition of energy, degrees of freedom and molecular basis of heat capacities.	

Course Learning Outcomes

1. Knowledge of evolution of scientific theories to explain the atomic structure, molecular geometry and physico-chemical behaviour of atomic matter made from elements in periodic table.
2. Focus on fundamentals of organic molecules, structure, stereochemistry, bonding, reactivity and reaction mechanisms.
3. Familiarization with gaseous state of matter and its physical laws governing it

List of Experiments

Inorganic Chemistry Practicals

1. Titrimetric Analysis
 - a. Calibration and use of apparatus.
 - b. Preparation of solutions of different Molarity/Normality of titrants.
 - c. Use of primary and secondary standard solutions.
2. Acid-Base Titrations
 - a. Estimation of carbonate and hydroxide present together in mixture.
 - b. Estimation of carbonate and bicarbonate present together in a mixture.

Organic Chemistry Practicals

3. Chromatography
 - a. Separation of a mixture of two amino acids by ascending and horizontal paper chromatography
 - b. Separation of a mixture of two sugars by ascending paper chromatography
 - c. Separation of a mixture of o-and p-nitrophenol or o-and p-aminophenol by thin layer chromatography (TLC).

Physical Chemistry Practicals

4. Surface tension measurements
 - a. Determine the surface tension by (i) drop number (ii) drop weight method.
 - b. Study the variation of surface tension of detergent solutions with concentration.
5. Viscosity measurements using Ostwald's viscometer

Determine of viscosity of aqueous solutions of (i) ethanol (ii) sugar at room temperature.

Text Books

A. Theory

Author	Title	Publisher	Ed/year	ISBN No	Pages
J.D. Lee	Concise Inorganic Chemistry	John Wiley and Sons Ltd	5th edition (2016)	978-8126515547	---
Greenwood, Earnshaw	Chemistry of the Elements	Butterworth - Heinemann	2nd edition (1997)	978-0750633659	----
M. B. Smith, J. March, March's	Advanced Organic Chemistry: Reactions, Mechanisms, and Structure	Wiley-Interscience	8th Edition (2015)	978-8126556588	---
Atkins P.W, Julio de Paula	Physical Chemistry	Oxford University Press, ELBS	11th edition (2018)	978-0198814740	---

Practicals

Author	Title	Publisher	Ed/year	ISBN No	Pages
J. Mendham, R.C. Denney, J. D. Barnes, M.J.K. Thomas , Vogel's	Vogel's Quantitative Chemical Analysis	Longman	6th edition (1999)	978- 0582226289	---
A.I. Vogel, A.R. Tatchell, B.S. Furnis	Vogel's Textbook of Practical Organic Chemistry	Prentice Hall	5th edition (2003)	978- 0582462366	----
Shoemaker, D.P Garland, C.W Nibler, J.W.	Experiments in Physical Chemistry	McGraw Hill Inc	8th edition (2008)	978- 0070570078	---

MAT114: Statistics for Biosciences

L	T	P	TOTAL CREDIT UNITS
2	0	0	2

Course Contents/syllabus:

	Teaching Hours
Unit I:	9 hrs
Data collection and graphical presentation, Descriptive Statistics: Measures of central tendency-Arithmetic, geometric and harmonic mean, median, and mode.	
Unit II:	9 hrs
Measures of dispersion, Skewness and Kurtosis, Correlation, and regression	
Unit III:	9 hrs
Definitions of Probability, Conditional Probability, Bayes' theorem, random variables: discrete and continuous, density and mass functions.	
Unit IV:	9 hrs
Expected values and moment generating functions, Discrete distributions: Uniform, Bernoulli, Binomial, Poisson, Continuous distributions: Uniform and Normal distribution	

Course Learning Outcomes: On the successful completion of this course,

1. Students will understand the concept of data collection, representation, and measures of central tendency
2. Students will be able to apply the concept of dispersion, skewness, correlation, and regression of the given data
3. Students will be having knowledge of probability and random variables.
4. Students will be able to apply the concepts of probability and random variables to different distributions

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
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Ronald E. Walpole, Raymond H. Myers, Sharon L. Myers, Keying E. Ye	Probability and Statistics for Engineers and Scientists	Pearson; 9th edition	2010	978-0321629111
G Shanker Rao	Probability and Statistics for Science and Engineering	Universities Press	2011	9788173717444
SC Gupta, VK Kapoor	Fundamentals of Mathematical Statistics	Sultan Chand & Sons Private Limited	2000	9788180545283

ENV106: Environmental Studies-II

L	T	P	Total Credits
2	0	0	2

Course content and syllabus

	Teaching Hours
Unit I: Environmental Pollution	9 hrs
<i>Environmental Pollution: types, Cause, effects and controls –Air, water, soil, chemical and noise pollution.</i> Nuclear hazard and human health risk Solid waste Management-control measures of urban and industrial waste. Pollution case studies.	
Unit II: Environmental Policies and Practices	9 hrs
<i>Environmental Policies and practices:</i> Climate change, global warming, ozone layer depletion, acid rain and impacts on human communities and agriculture. Environment laws: Environment Protection Act; Air (Prevention and Control of Pollution) Act; Water (Prevention and Control of Pollution) Act; Wildlife Protection Act; Forest Conservation Act, international agreements: Montreal and Kyoto protocols and convention on biological diversity(CBD), The Chemical Weapons Convention (CWC). Natural reserves, tribal population and rights and Human-wildlife conflict in Indian context.	
Unit III: Human communities and the environment	9 hrs

<p>Impacts on environment, human health and welfare. Carbon foot-print.</p> <p>Resettlements and rehabilitation of project affected persons, case studies. Disaster management: floods, earthquake, cyclone and landslides.</p> <p>Environmental movements: Chipko, Silent valley, Bishnois of Rajasthan. Environmental ethics: Role of Indian and other religions and cultures in environmental conservation.</p> <p>Environmental communication and public awareness, case studies (e.g., CNG vehicles in Delhi).</p>	
Unit IV: Field Work	9 hrs
<ul style="list-style-type: none"> • Visit to an area to document environmental assets: river/forest/flora/fauna, etc. • Visit to local polluted Site-Urban/Rural/Industrial/Agricultural • Study of common plants, insects, birds and basic principles of identification. • Study of simple ecosystems-pond, river, Delhi Ridge, etc 	

Course Learning Outcomes:

1. Understanding the types of pollution and their impact on environment and human health.
2. Understand the environmental concerns and their impact on humans and agriculture.
3. Able to analyse the impacts of natural and manmade disaster on human population and settlements.
4. Sensitization about the environmental issues and concerns leading to proactive actions to improve the environmental conditions in our daily life.
5. Able to imbibe practical approach and solution to solve environmental concerns.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
William P. Cunningham , Mary Ann Cunningham	Principles of Environmental Science	McGraw-Hill	2019	9781260219715	464
William P. Cunningham , Mary Ann Cunningham , Barbara	Environmental Science: A global concern	McGraw-Hill	2021	9781260363821	624

Woodworth Saigo					
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ENG103: Communication Skills—II

L	T	P/S	SW/FW	TOTAL CREDIT UNITS
1	0	0	0	1

Course Contents/syllabus:

	Teaching Hrs (H)
Unit I: Basic Concepts in Communication	5 hrs
Towards communicative competence; choosing the appropriate channel and medium of communication; ways to develop communication skills in the areas of Listening, Speaking, Reading, and Writing.	
Unit II: Communication Types	4 hrs
Nonverbal communication: detailed analysis, KOPPACT (Kinesics, Oculistics, Proxemics, Paralanguage, Artefacts, Chronemics, Tactilics).	
Unit III: Communication and Technology	5 hrs
Importance of digital literacy and communication on digital platforms.	
Unit IV: Presentation Skills	5 hrs
Planning, preparation, practice, and performance; audience analysis, audio-visual aids, analyzing the non-verbal communication, methods of delivery: impromptu, extemporaneous, memorization, manuscript, and outlining.	

Course Learning Outcomes:

1. Students will be able to understand the need and the methods required to develop communication skills in the areas of listening, speaking, reading, and writing.
2. Students will be able to understand the significance of non-verbal communication in various contexts.
3. Students will be able to develop an awareness of the role of digital platforms in shaping public psyche, beliefs, and perceptions about social realities and build an informed and critical perspective.
4. Students will be able to develop and upgrade their presentation skills.

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
P. D. Chaturvedi and Mukesh Chaturvedi	Business Communication: Concepts, Cases and Applications	Pearson Education	2006	9788131701720
Meenakshi Ramanand Prakash Singh	Business Communication	Oxford University Press	2012	9780198077053
Jeff Butterfield	Soft Skills for Everyone	Cengage Learning	2017	9789353501051

PSY106: INDIVIDUAL, SOCIETY AND NATIO

L	T	P/S	SW/FW / PSDA	TOTAL CREDIT UNITS
1	0	0	0	1

Course Contents/syllabus:

	Teaching Hrs
Unit-1- Individual differences & Personality	4 hrs
<ul style="list-style-type: none"> • Personality: Definition & Relevance • Importance of nature & nurture in Personality Development • Importance and Recognition of Individual differences in Personality • Accepting and Managing Individual differences Intuition, Judgment, Perception & Sensation (MBTI) BIG5 Factors 	
Unit-2- Managing Diversity	5 hrs
<ul style="list-style-type: none"> • Defining Diversity • Affirmation Action and Managing Diversity • Increasing Diversity in Work Force • Barriers and Challenges in Managing Diversity 	
Unit-3- Socialization, Patriotism and National Pride	4 hrs
<ul style="list-style-type: none"> • Nature of Socialization • Social Interaction • Interaction of Socialization Process • Contributions to Society and Nation • Sense of pride and patriotism • Importance of discipline and hard work • Integrity and accountability 	
Unit-4- Human Rights, Values and Ethics	5 hrs

<ul style="list-style-type: none"> • Meaning and Importance of human rights • Human rights awareness • Values and Ethics- Learning based on project work on Scriptures like- Ramayana, Mahabharata, Gita etc. 	
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List of Professional Skill Development Activities (PSDA):

1. Project on Understanding Diversity
2. Term Paper on Patriotism among Youth

Course Learning Outcomes: On completion of the course:

1. To recognize individual differences
2. To manage individual differences
3. To develop patriotic feelings
4. To recognized their self in relation to society & nation

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN	Pages
Department of English, University of Delhi	The Individual & Society	Pearson Education	2010	978-8131704172	266
Umang Malhotra	Individual, Society, and the World	iUniverse	2004	978-0595662401	188
Tonja R. Conerly & Kathleen Holmes	Introduction to Sociology 3e	Openstax	2015	9781711493978	458
Daksh Tyagi	“A Nation of Idiots”	Every Protest	2019	978-8194275015	350

FOL103: French Grammar

L	T	P	Total Credits
1	0	0	1

Course content and syllabus

	Teaching Hours
Unit I: My family and my house	5 hrs
Descriptors/Topics <ul style="list-style-type: none">• Talk about your family members• Usage of possessive adjectives• Describe your house/apartment• Prepositions of location• Negation	
Unit II: Lifestyle	4 hrs
Descriptors/Topics <ul style="list-style-type: none">• Talk about your hobbies and pastimes• Usage of appropriate articles : definite and contracted• Talk about your daily routine• Usage of pronominal verbs	
Unit III: In the city	5 hrs
Descriptors/Topics <ul style="list-style-type: none">• Filling up a simple form• Ask for personal information• Usage of interrogative adjectives• Give directions about a place	

<ul style="list-style-type: none"> • Ordinal numbers • Usage of demonstrative adjectives 	
Unit IV: Week-end	4 hrs
Descriptors/Topics <ul style="list-style-type: none"> • Talk about your week-end plans • Usage of disjunctive pronouns • Usage of Near Future tense • Talk about weather • Write a simple post card 	

Course Learning Outcomes: At the end of this course, the students will be able to interact in a simple way on everyday topics. This course content focuses on the speech of the students in a lucid and a concurrent manner using appropriate vocabulary and pronunciation techniques. Extra stress will be given on their understanding of grammatical structures and the foreign accent of the language. At the end of the course, the student shall be able to:

1. Understand information; Express in his own words; Paraphrase; Interpret and translate.
2. Apply information in a new way in a practical context
3. Analyze and break-down information to create new ideas
4. Evaluate and express opinion in a given context

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Christine Andant, Catherine Metton, Annabelle Nachon, Fabienne Nugue	A Propos - A1, Livre de l'élève et Cahier d'exercices.	Langers International Pvt. Ltd.	2010	9789380809069	---
Collins Dictionaries	Easy Learning French Complete Grammar,	Collins	2016	978-0008141721	---

	Verbs and Vocabulary				
Nikita Desai, Samapita Dey Sarkar	Apprenons La Grammaire Ensemble - French	Langers Internation alPvt. Ltd.	2017	978- 8193002681	---

FOL104: German Grammar

L	T	P	Total Credits
1	0	0	1

Course content and syllabus

	Teaching Hours
Unit I: Time (Uhrzeit); People and the World: Land, Nationalität und Sprache	4 hrs
<ul style="list-style-type: none"> • Introduction of time • Read text related to time and teach the students the time expressions • Exercises related to Time • Adverbs of time and time related prepositions • Vocabulary: Countries, Nationalities, and their languages • Negation: "nicht/ kein" • Ja/Nein Fragen. • All the colors and color related vocabulary, adjectives, and opposites • Exercises and comprehension for the same 	
Unit II: Irregular verbs (unregelmässige Verben)	5 hrs
<ul style="list-style-type: none"> • Introduction to irregular verbs and their conjugation e.g. fahren, essen, lesen etc • Read a text related to the eating habits of Germans • Vocabulary: Obst, Gemüse, Kleiderstück with usage of irregular verbs • Free time and hobbies • Food and drinks 	
Unit III: Accusative case: articles and pronouns (Akkusativ Kasus: Artikel und Pronomen)	5 hrs

<ul style="list-style-type: none"> • Introduction to the concept of object (Akkusativ) • Formation of sentences along with the translation and difference between nominative and accusative articles • Usage of accusative Definite articles • Usage of accusative Indefinite articles 	
Unit IV: Accusative case: possessive pronouns (Akkusativ Kasus: Possessivpronomen) Family and Relationship	4 hrs
<ul style="list-style-type: none"> • Accusative Personal Pronouns: - Revision of the nominative personal pronouns and introduction of accusative. Applicability of pronouns for both persons and things. • Usage of accusative Personal Pronouns • Introduction of accusative possessive pronouns • Difference between nominative and accusative possessive pronouns • usage of accusative possessive pronouns 	

Course Learning Outcomes: After completing these modules, the students will be capable of constructing sentences with possessive and demonstrative adjectives in German. In addition, they will be proficient in formulating meaningful sentences as they will be capable of applying their knowledge of all the irregular verbs they have learnt during the session. They will also have an idea of German culture by studying about various German festivals.

At the end of the course, the student shall be able to:

1. Understand information; Express in his own words; Paraphrase; Interpret and translate.
2. Apply information in a new way in a practical context
3. Analyse and break-down information to create new ideas
4. Evaluate and express opinion in a given context

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Dora Schulz, Heinz Griesbach	Deutsche Sprachlehre Fur Auslander	Max Hueber Verlag	1984	978-3190010066	---

Hartmut Aufderstrasse, Jutta Muller, Helmut Muller	Themen Aktuell: Glossar Deutsch	Max Hueber Verlag	2003	978-3190816903	---
Giorgio Motta	Wir Plus Grundkurs Deutsch für Junge Lerner Book German Guide	Goyal Publishers	2011		248

INL104: Punjabi Language and Literature

L	T	P	Total Credits
1	0	0	1

Course content and syllabus

	Teaching Hours
Unit I:	4 hours
<p>ਆਧੁਨਿਕ ਪੰਜਾਬੀ ਕਹਾਣੀ ਦਾ ਅਧਿਐਨ (ਕਥਾ ਕਹਾਣੀ) ਕਹਾਣੀ ਵਿਸ਼ਾ-ਵਸਤੂ/ਸਾਰ,ਪਾਤਰ-ਚਿਤਰਨ ਕਹਾਣੀਕਾਰ ਦੇ ਜੀਵਨ ਅਤੇ ਰਚਨਾ ਬਾਰੇ ਮੁੱਢਲੀ ਜਾਣਕਾਰੀ</p>	
Unit II:	4 hours
<p>ਦਫ਼ਤਰੀ ਚਿੱਠੀ-ਪੱਤਰ ਰਚਨਾ ਚਿੱਠੀ-ਪੱਤਰ ਲੇਖਣ ਕਲਾ,ਮਹੱਤਤਾ ਅਤੇ ਕਿਸਮਾਂ ਦਫ਼ਤਰੀ ਚਿੱਠੀ-ਪੱਤਰ ਰਚਨਾ ਦੇ ਜ਼ਰੂਰੀ ਅੰਗ ਅਤੇ ਵੱਖ-ਵੱਖ ਵਿਸ਼ਿਆਂ ਅਨੁਸਾਰ ਵਿਹਾਰਕ ਅਭਿਆਸ</p>	
Unit III:	5 hours
<p>ਵਿਆਕਰਨ ਸਿਧਾਂਤ ਅਤੇ ਵਿਹਾਰ</p> <p>1. ਪੰਜਾਬੀ ਅਰਥ ਬੋਧ ਅਰਥਾਂ ਦੇ ਆਧਾਰ ਦੇ ਸ਼ਬਦਾਂ ਦੀਆਂ ਕਿਸਮਾਂ ਅਤੇ ਉਦਾਹਰਨਾਂ, ਸਮਾਨਰਥਕ ਸ਼ਬਦ, ਬਹੁਅਰਥਕ ਸ਼ਬਦ, ਵਿਰੋਧਾਰਥਕ ਸ਼ਬਦ, ਬਹੁਤੇ ਸ਼ਬਦਾਂ ਦੇ ਸਥਾਨ ਤੇ ਇੱਕ ਸ਼ਬਦ ਮੁਹਾਵਰੇ, ਅਖਾਣ : ਪਰਿਭਾਸ਼ਾ ਅਤੇ ਉਦਾਹਰਨਾਂ</p> <p>2. ਪੰਜਾਬੀ ਵਾਕ ਬੋਧ ਵਾਕ ਪ੍ਰੀਭਾਸ਼ਾ,ਵਾਕ ਦੇ ਤੱਤ, ਪੰਜਾਬੀ ਵਾਕ ਤਰਤੀਬ ਵਾਕ ਵਰਗੀਕਰਨ:ਕਾਰਜ ਦੇ ਅਧਾਰ ਤੇ ਵਾਕਾਂ ਦੀਆਂ ਕਿਸਮਾਂ, ਬਣਤਰ ਦੇ ਅਧਾਰ ਤੇ ਵਾਕਾਂ ਦੀਆਂ ਕਿਸਮਾਂ</p>	
Unit IV:	5 hours
<p>ਵਿਆਕਰਨ ਸਿਧਾਂਤ ਅਤੇ ਵਿਹਾਰ</p> <p>1. ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਅਤੇ ਗੁਰਮੁਖੀ ਲਿੱਪੀ</p>	

2. ਭਾਸ਼ਾ, ਉਪਭਾਸ਼ਾ, ਟਕਸਾਲੀ ਭਾਸ਼ਾ ਅਤੇ ਪੰਜਾਬੀ ਦੀਆਂ ਉਪਭਾਸ਼ਾਵਾਂ	
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Course Learning Outcomes:

1. Understand modern Punjabi Stories.
2. Interpret the importance of letter writing
3. Analyze the Punjabi language structure and grammar.
4. Examine the impact and importance of Punjabi dialects and Gurmukhi script on Punjabi language.

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN	Pages
ਡਾ. ਧਨਵੰਤ ਕੋਰ (ਸੰਪਾ.),	ਕਥਾ ਕਹਾਣੀ	ਪਬਲੀਕੇਸ਼ਨ ਬਿਊਰੋ, ਪੰਜਾਬ ਯੂਨੀਵਰਸਿਟੀ ਚੰਡੀਗੜ੍ਹ	2009	-	-
ਸੁਰਿੰਦਰ ਸਿੰਘ ਖਹਿਰਾ (ਸੰਪਾ.),	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਵਿਆਕਰਨ ਅਤੇ ਬਣਤਰ	ਪਬਲੀਕੇਸ਼ਨ ਬਿਊਰੋ, ਪੰਜਾਬੀ ਯੂਨੀਵਰਸਿਟੀ ਪਟਿਆਲਾ	2015	-	-
ਡਾ. ਹਰਕੀਰਤ ਸਿੰਘ,	ਕਾਲਜ ਪੰਜਾਬੀ ਵਿਆਕਰਨ ਅਤੇ ਲੇਖ ਰਚਨਾ	ਪੰਜਾਬ ਸਟੇਟ ਯੂਨੀਵਰਸਿਟੀ ਟੈਕਸਟ ਬੁੱਕ ਬੋਰਡ, ਚੰਡੀਗੜ੍ਹ	1999	-	-
ਡਾ. ਪ੍ਰੇਮ ਪ੍ਰਕਾਸ਼ ਸਿੰਘ	ਕਾਲਜ ਪੰਜਾਬੀ ਵਿਆਕਰਨ ਅਤੇ ਲੇਖ ਰਚਨਾ	ਮਦਾਨ ਪਬਲੀਕੇਸ਼ਨਜ਼, ਪਟਿਆਲਾ	2002	-	-
ਡਾ. ਬੂਟਾ ਸਿੰਘ ਬਰਾੜ	ਪੰਜਾਬੀ ਵਿਆਕਰਨ ਸਿਧਾਂਤ ਅਤੇ ਵਿਹਾਰ	ਚੇਤਨਾ ਪ੍ਰਕਾਸ਼ਨ, ਪੰਜਾਬੀ ਭਵਨ, ਲੁਧਿਆਣਾ	2012	-	-
ਡਾ. ਬੂਟਾ ਸਿੰਘ ਬਰਾੜ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਸ਼੍ਰੋਤ ਅਤੇ ਸਰੂਪ	, ਵਾਰਿਸ ਸ਼ਾਹ ਫ਼ਾਊਂਡੇਸ਼ਨ, ਅੰਮ੍ਰਿਤਸਰ	2012	-	-
ਦੁਨੀ ਚੰਦ੍ਰ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਦਾ ਵਿਆਕਰਣ	, ਪੰਜਾਬ ਯੂਨੀਵਰਸਿਟੀ ਪਬਲੀਕੇਸ਼ਨ ਬਿਊਰੋ, ਚੰਡੀਗੜ੍ਹ	1995	-	-

ਜੋਗਿੰਦਰ ਸਿੰਘ ਪੁਆਰ ਅਤੇ ਹੋਰ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਦਾ ਵਿਆਕਰਨ (ਭਾਗ 1,2,3),	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਅਕਾਦਮੀ ਜਲੰਧਰ	2003	-	-
ਸੁਖਵਿੰਦਰ ਸਿੰਘ ਸੰਘਾ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਵਿਗਿਆਨ	ਪੰਜਾਬੀ ਭਾਸ਼ਾ ਅਕਾਦਮੀ ਜਲੰਧਰ	2010	--	-
ਅਗਨੀਹੋਤਰੀ, ਵੇਦ	ਪਰਿਚਾਇਕ ਭਾਸ਼ਾ ਵਿਗਿਆਨ	ਦੀਪਕ ਪਬਲਿਸ਼ਰਜ਼ ਜਲੰਧਰ	1981	-	-

INL106: History and Culture of Punjab

L	T	P/S	SW/F W	Total Credit Units
1	0	0	0	1

Course Contents/syllabus:

	Teaching Hrs
Unit I:	4H
1. Introduction of Colonial Rule in Punjab: Annexation of Punjab; Board of Administration. 2. Western Education: Growth of Education and rise of middle classes. 3. Agrarian Development: Commercialization of agriculture; canalization and colonization.	
Unit II:	5H
4. Early Socio Religious Reform: Christian Missionaries; Namdharis; Nirankaris. 5. Socio Religious Reform Movements: activities of Arya Samaj; Singh sabhas; Ahmadiyahs; Ad Dharam Movement 6. Development of Press & literature: growth of print technology; development in literature	
Unit III:	4H
7. Emergence of Political Consciousness: Gadar Movement; Jallianwala Bagh Massacre 8. Gurudwara Reform Movement; major Morchas; Activities of Babbar Akalis. 9. Struggle for Freedom: Non-Cooperation Movement; HSRA and Bhagat Singh; Civil Disobedience Movement; Quit India Movement.	
Unit IV:	4H
10. Partition and its Aftermath: resettlement; rehabilitation 11. Post-Independence Punjab: Linguistic Reorganization; Green Revolution.	

Course Learning Outcomes:

1. Understand the history of Punjab region in modern times.
2. Interpret the importance early socio religious reform, movements, developments.

3. Examine the contribution of major reform movements: Gadar, Babbar Akalis and Gurdwarareform morchas.
4. Examine the impact of Partition of Punjab and major changes in Punjab after independence.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Singh, Kirpal	History and Culture of the Punjab, Part II (Medieval Period)	Publication Bureau, Punjabi University, Patiala	1990(3rd ed.).	---	---
Singh, Fauja(ed.)	History of the Punjab, Vol.III	Punjabi University, Patiala	1972	---	---
Grewal, J.S.	The Sikhs of the Punjab, the New Cambridge History of India	Orient Longman	1990	---	---
Singh, Khushwant	A History of the Sikhs, vol I: 1469-1839	oxford University Press, Delhi	1991	---	---
Chopra, P.N., Puri, B.N.	A Social, Cultural and Economic History of India, Vol.II, And Das	M.N. Macmillan , Delhi	1974	---	---

Programme structure for B.Sc. (H) Microbiology- 3 years (3rd Semester)

Sr. No	Course Code	Course Title	Course Type	Credits					Credit Units
				L	T	P	FW	SW	
1	BTY201	Biotechniques	Core Course	4	0	2	0	0	6
2	BCH201	Enzymology	Core Course	4	0	2	0	0	6
3	MBO201	Mycology and Phycology	Core Course	4	0	2	0	0	6
4	BTY202	Protein Science	Allied Course	4	0	0	0	0	0
5	CAS211	Programming in C	Skill Enhancement Course	1	0	1	0	0	2

Total Credits

24

BTY201: Biotechniques

L	T	P	Total Credits
4	0	2	6

Course Objective: This course will provide students with the understanding of various analytical techniques used in biology/biotechnology-based research and industry. The course will acquaint the students with the various instruments, their configuration and principle of working, operating procedures, data generation and its analysis.

Course content and syllabus

	Teaching Hours
Unit I: Introduction to chromatographic techniques	18 hrs
Theoretical basis of chromatographic separations, Principles and applications of paper, thin layer, column, ion-exchange, affinity, gel permeation, normal phase and reverse phase chromatography, gas chromatography, High performance liquid chromatography (HPLC)	
Unit II: Spectroscopic and centrifugation techniques	18 hrs
Theory and application of UV-Visible, Infrared, Raman, Nuclear magnetic resonance, Fluorescence, Atomic absorption spectroscopy, X-ray diffraction, Introduction to mass spectroscopy, Introduction to centrifugation, basic principles of sedimentation, types of centrifuges and their uses, density gradient and analytical centrifugation, sub-cellular fractionation	
Unit III: Electrophoretic techniques	18 hrs
Theoretical basis of electrophoretic separations, electrophoretic mobility, moving boundary electrophoresis, paper, starch-gel, polyacrylamide gel (native and SDS-PAGE), agarose-gel electrophoresis, pulse-field gel electrophoresis, immune-electrophoresis, isoelectric focusing, western blotting	
Unit IV: Microscopy	18 hrs
Simple microscopy, phase contrast microscopy, fluorescence, and electron microscopy (Transmission and Scanning)	

List of Experiments:

1. To verify the validity of Beer's law and determine the molar extinction coefficient of KMNO₄ and K₂Cr₂O₇
2. Separation of amino acids by paper chromatography
3. To identify biomolecules in a given sample by paper/thin layer chromatography
4. Demonstration of HPLC instrument
5. Determination of concentration of metal ion using atomic absorption spectroscopy
6. Demonstration of working of centrifuge
7. Preparation of sub-cellular fractions of cells
8. Native and SDS-polyacrylamide gel electrophoresis of proteins.

Course Learning Outcomes:

Students will be able to

1. apply basic principles of different analytical techniques in analytical work.
2. use spectroscopy and chromatography in biotechnological applications.
3. use microscopy, centrifugation, and electrophoretic techniques.
4. demonstrate principle and working of various instruments.
5. use various techniques for solving industrial and research problems.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Karp, Gerald	Cell and Molecular Biology: Concepts and Experiments	John Wiley and Sons, Inc	6th edition/2010	111830179X 978- 1118301791	783
Wilson K., Walker J.	Principle and Techniques of Biochemistry and Molecular Biology	Cambridge University Press	6th edition/2006	0521178746 9780521178747	744
Plummer, David	An Introduction to Practical Biochemistry	Tata Mc GrawHills	3rd edition/2017	0070994870 9780070994874	332

BCH201: Enzymology

L	T	P	Total Credits
4	0	2	6

Course Objectives: Explore enzymology to understand enzyme structure, function, and kinetics, facilitating insights into biological processes, industrial applications, and the development of therapeutic interventions.

Content and syllabus

	Teaching Hours
Unit I: Enzymes and Coenzymes	18 hrs
Nature of enzymes - protein and non-protein (ribozyme). Cofactor and prosthetic group, apoenzyme, holoenzyme. IUBMB classification of enzymes. Features of enzyme catalysis Factors affecting the rate of chemical reactions, collision theory, activation energy and transition state theory, catalysis, reaction rates and thermodynamics of reaction. Catalytic power and specificity of enzymes (concept of active site), Koshland's induced fit hypothesis. Involvement of coenzymes in enzyme catalysed reactions: Mechanism of action of TPP, FAD, NAD, pyridoxal phosphate, biotin, coenzyme A, tetrahydrofolate, lipoic acid.	
Unit II: Enzyme Kinetics and Inhibition	18 hrs
Relationship between initial velocity and substrate concentration, steady state kinetics, equilibrium constant – mono-substrate reactions. Michaelis-Menten equation, Lineweaver-Burk plot, Eadie-Hofstee and Hanes plot. Km and Vmax, Kcat and turnover number. Effect of pH, temperature and metal ions on the activity of enzyme. Bi-substrate reactions: Types of bi bi reactions (sequential – ordered and random, ping pong reactions). Enzyme inhibition: Reversible inhibition and irreversible (competitive, uncompetitive, noncompetitive, mixed type). Mechanism based inhibitors - antibiotics as inhibitors.	
Unit III: Mechanisms of Enzyme catalysed reactions	18 hrs

General features - proximity and orientation, strain and distortion, acid base and covalent catalysis (chymotrypsin, lysozyme). Metal activated enzymes and metalloenzymes, transition state analogues. Regulation of enzyme activity : Control of activities of enzymes (end product inhibition) and metabolic pathways, feedback inhibition (aspartate transcarbamoylase), reversible	
covalent modification (phosphorylation). Proteolytic cleavage- zymogen. Multienzyme complexes (pyruvate dehydrogenase, fatty acid synthase) and Enzyme regulation	
Unit IV: Application of Enzymes	18 hrs
Application of enzymes in diagnostics (SGPT, SGOT, creatine kinase, alkaline and acid phosphatases), enzyme immunoassay (HRPO), enzyme therapy (Streptokinase). Immobilized enzymes. Isoenzymes Enzyme Inhibitors as drugs. Drug Design	

List of Experiments -with basic instructions (Total Teaching = 60 hrs)

1. Partial purification of acid phosphatase from germinating mung bean.
2. Assay of enzyme activity and specific activity, e.g. acid/alkaline phosphatase.
3. Effect of pH on enzyme activity
4. Effect of temperature on enzyme activity
4. Determination of K_m and V_{max} using Lineweaver-Burk plot
5. Enzyme inhibition - calculation of K_i for competitive inhibition.
6. Continuous assay of lactate dehydrogenase.
7. Coupled assay of glucose-6-phosphate dehydrogenase.

Course Learning Outcomes: at the end of the course, the students will learn

1. Types of enzymes, classification and their importance
2. Enzyme kinetics and enzyme inhibitors
3. Mechanisms of enzyme action
4. Application of enzymes in diagnostics and drug discovery

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
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David L. Nelson	Lehninger: Principles of Biochemistry	W. H. Freeman	2013	978-1429234146 1429234148	1328
Nicholas C.P. and Lewis S.	Fundamentals of Enzymology	Oxford University Press	1989	019806439X	526
				978-0198064398	
Voet, D., Voet, J.G.	Biochemistry	Wiley	2010	0121822117 978-0470570951	1428

MBO201: Mycology and Phycology

L	T	P	Total Credits
4	0	2	6

Course Objectives: Investigate mycology and phycology to comprehend the diversity, ecology, and significance of fungi and algae in various ecosystems, industries, and human health, fostering a deeper understanding of their roles and applications in biotechnology, agriculture, and environmental management.

Course content and syllabus

	Teaching Hours
Unit I: MYCOLOGY - PHYLOGENY AND SYSTEMATICS	18 hrs
General characteristics; Ecology and distribution, Classification and General account of major classes of Fungi; Habit, Structure and Methods of Reproduction in Basidiomycotina, Deuteromycotina and Mycoplasma. Occurrence, General structure, Nutrition, Reproduction, Fungi as infectious agents – plants, animals, and humans. Fear of fungi - extinction of species.	
Unit II: PHYCOLOGY - PHYLOGENY AND SYSTEMATICS	18 hrs
General characteristics; Ecology and distribution; range of thallus organization; Cell structure and components; cell wall, pigment system, reserve food, flagella; methods of reproduction; Classification; criteria, system of Fritsch, and evolutionary classification of Lee (only upto groups); Role of algae in the environment, agriculture, biotechnology and industry.	
Unit III: MOLECULAR MYCOLOGY AND PHYCOLOGY	18 hrs
Role of modern techniques like PCR, Chromatography and Spectroscopy in identification and characterization of species; Current advances in algal and fungal research; yeast and fungal genetics; yeast and fungi as genetic models to study human physiology, biochemistry, and disease.	
Unit IV: YEAST AND FUNGI AS INDUSTRIAL WORKHORSES	18 hrs
Role of yeast, fungi, and algae in food, pharmaceutical, and green chemistry industries.	

Secondary metabolites – production, biochemistry, and mechanisms of action.	
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List of Experiments -with basic instructions (Total Teaching = 60 hrs)

1. Identification and characterization of species by microscopy.
2. Isolation and purification of microorganisms from soil/water/air by streak plate method and serial dilution.
3. To perform the lactophenol blue staining of the purified fungal culture.
4. To test for the antibiotic sensitivity of the yeast and fungal sample.
5. To perform the MIC test for antibiotic sensitivity of a yeast and fungal strain against a specific antibiotic.
6. To perform standard growth curve of purified yeast and fungal strain.
7. To study the effect of temperature and pH on the growth of microorganisms.
8. Preservation of a microbial strain.

Course Learning Outcomes:

1. Understand Phycology and mycology – occurrence and ecology.
2. Evaluate bacterial classification and diversity
3. Role of mycology in bio-medical research
4. Perceive knowledge of applications of yeast, fungi and algae.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Bridge, P.D. CAB Intern. 1993..	Application of PCR in Mycology.	Ferry Lane, Kew, U.K.	1998		
Cook, R. C. and M. J. Whipps.	Ecophysiology of Fungi.	Blackwell Scientific, London, UK	1993	0632021683 978063202168 0	337

Pelczar, M.J. Jr., Chan ECS and Krieg, N.R.	Microbiology: Concepts and Applications	New York; Madrid : McGraw-Hill,	1993	0070492581, 978007049258 5	
Cappucino, J.G.	Microbiology-A laboratory manual, 4th ed., Harlow, Addition- Wesley.	Hoboken, N.J.: Pearson	2020	0135188997, 978013520399 6, 0135203996	

BTY202: Protein Science

L	T	P	Total Credits
4	0	0	4

Course Objectives: Explore protein science to understand protein structure, function, and interactions, enabling the analysis of biological processes, drug discovery, and the development of biotechnological applications

Course content and syllabus

	Teaching Hours
Unit I: Protein Structure	18 hrs
Peptide bond, protein secondary structure – fibrous and globular proteins, proteins stability, tertiary and quaternary structure, Protein Folding: Theory and Experiment, Folding Accessory Proteins, Protein Structure Prediction and Design, Protein Dynamics.	
Unit II: Protein misfolding, aggregation and denaturation	18 hrs
Protein misfolding and aggregation, amyloid formation. Conformational Diseases: Alzheimer's, Prion diseases, Huntington's disease, sickle cell anemia, Parkinsons. Structural Evolution Protein denaturation and folding, Chemical evolution, Chemical Synthesis of Polypeptides. IDP (Intrinsically disordered proteins).	
Unit III: Protein alignment and database research	18 hrs
Protein primary sequence analysis, DNA sequence analysis, pair wise sequence alignment, FASTA algorithm, BLAST, multiple sequence alignment, DATA base searching using BLAST and FASTA. Phylogenetic tree analysis	
Unit IV: Analysis of protein-protein interactions-	18 hrs
Pull-down assay, Yeast two hybrid assay, Coimmunoprecipitation assay, Fluorescenceresonance energy transfer (FRET). DNA- protein interactions, footprinting assay, EMSA.	

Course Learning Outcomes:

1. Understand basic concepts of protein structure.
2. Describe and understand the thermodynamic stability of proteins. Be able to apply this knowledge to interpret experimental data and calculate thermodynamic parameters for proteins with different mechanisms of folding.

3. Describe the energy landscape of protein folding and misfolding from a thermodynamic point of view.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Donald Voet, Judith G. Voet	Biochemistry, 4th Edition	John Wiley & Sons	2015	ISBN: 978-0- 470-57095-1	1520
David L. Nelson and Michael M. Cox	Lehninger Principles of biochemistry, 8 th Edition	Macmillan	2021	ISBN:9781319 322328	1120

CAS211: Programming with C

L	T	P	SW/FW	No. of PSDA	TOTAL CREDIT UNITS
1	0	1	0	0	2

Course Objectives: The course is designed to provide basic knowledge of procedural programming and learn constructs of C language. Students will be able to develop logics which will help them to create programs in C.

Course Contents/syllabus:

	Teaching Hours
Unit I: Introduction of Programming Languages	5 H
Introduction: Types of Languages, Evolution of 'C' Language, Structure of a 'C' Program, 'C' Program development life cycle, Executing and Debugging a 'C' Program. 'C' Tokens: Keywords and Identifiers, Operators, Constants, Variables, Data Types, Precedence of Operators, Scope and Lifetime of Variables.	
Unit II: Control Statement and Looping	4 H
Control Statements: Decision Making using if statement, Types of if ...else block, Switchcase Block, Arithmetic Expressions, Evaluation of Expressions, GOTO statement Looping: Concept of Loop, For loop, While loop, Do while loop, Jumping in Loop, break and continue statement.	
Unit III: Arrays and Strings	4 H
Arrays and Strings: Introduction to array, Processing Array Contents, 2D arrays, Array with three or more dimensions. String, string concatenation, Comparing strings, String handling Functions.	
Unit IV: Functions, Structure and Unions	5 H

Function: Concept of Function, User defined Function, System Defined Function, Function Calling, Types of parameters passing in function, return type in Function.	
Structure & Union: Need of Structure, Implementing Structure Variable, Arrays of Structure, Structure within Structure, Introduction of Unions, Difference between Structure and Unions.	

Course Learning Outcomes: After studying this course students will be able:

1. To understand the fundamentals and tokens of C programming.
2. To develop skills to implement decision making through control structures in C.
3. To Analyze the working and implementation of array in memory.
4. To Optimize the code with the help of functions and structures.

Lab/ Practical details, if applicable:

Objective: The aim of this section of Lab is to teach experiments of C programming pertaining to the units beingtaught in the theory paper specifically related to procedural programming, strings, structures and unions.

1. Write a Program to read radius value from the keyboard and calculate the area of circle and print the result in both floating and exponential notation.
2. Write a Program to convert temperature. (Fahrenheit –Centigrade and vice-versa)
3. Write a program for computing the volume of sphere, cone and cylinder assume that dimensions are integer’s use type casting where ever necessary.
4. Write a Program to read marks of a student in six subjects and print whether pass or fail (using if-else).
5. Write a Program to calculate roots of quadratic equation (using if-else).
6. Write a Program to calculate electricity bill. Read starting and ending meter reading. The charges areas follows.

No. of Units Consumed	Rate in(Rs)
1-100	1.50 per unit
101-300	2.00 per unit for excess of 100 units
301-500	2.50 per unit for excess of 300 units
501-above	3.25 per unit for excess of 500 units

Do the Following Programs Using for, while, do-while loops.

7. Write a program to calculate sum of individual digits of a given number.
8. Write a program to check whether given number is palindrome or not.

9. Write a program to check whether a given number is a Fibonacci number or not.
10. Write a program to read 2 numbers x and n then compute the sum of the Geometric Progression. $1+x+x^2+x^3+\dots+x^n$
11. Write a program to print the following formats.

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1                *
1 2              * * *
1 2 3            * * * * *
1 2 3 4          * * * * * * *

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12. Write a program to perform matrix addition, matrix subtraction and transpose of a matrix.
13. Write a program to verify the given string is palindrome or not (without built-in functions, with using built-in functions).
14. Write a program to swap two numbers using a) Call By Value B) Call By Reference.
15. Write a program to create structure for an account holder in a bank with following Fields: name, account number, address, balance and display the details of five account holders.

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
Jeri R. Hanly, Elliot B. Koffman	Problem Solving and Program Design in C	Pearson	2015	978-0134014890
Pradip Dey, Manas Ghosh	Programming In C	Oxford University Press	2018	978-0199491476
E Balagurusamy	Programming in ANSIC	McGraw Hill Education	2019	978-9351343202
Yashwant Kanetkar	Let Us C	BPB Publications	2020	978-9389845686

Programme structure for B.Sc. (H) Microbiology- 3 years (4th Semester)

Sr . N o	Course Code	Course Title	Course Type	Credits					Credit Units
				L	T	P	FW	SW	
1	IMM202	Immunology	Core Course	4	0	2	0	0	6
2	BTY206	Bioprocess Technology	Core Course	4	0	2	0	0	6
3	MBO203	Microbial Physiology and Metabolism	Core Course	4	0	2	0	0	6
4	BTY207	Recombinant DNA Technology	Allied Course	3	0	1	0	0	4
5	PHY213	Fundamentals of Physics	Skill Enhancement Course	2	0	0	0	0	2

Total Credits

24

*The Skill Enhancement Course can also be taken through MOOC

IMM206: Immunology

L	T	P	Total Credits
4	0	2	6

Course Objective: The objective of this course is to provide students with detail understanding of different cells of the immune system and their role in immune protection as well as application of immunological techniques

Course content and syllabus

	Teaching Hours
Unit I: Introduction and Immune Cell Types	18 hrs
Immune system, Concept of Innate and Adaptive immunity, Hematopoietic stem cells, Lymphocytes & immune response (cytotoxic T-cell, helper T-cell, suppressor T-cells), Granulocytes and Monocytes, Cell participation in innate and adaptive Immunity	
Unit II: Antigens, Antibodies and Major Histocompatibility Complex	18 hrs
Characteristics of an antigen (Foreignness, Molecular size, and Heterogeneity), Haptens, Epitopes (T & B cell epitopes), T-dependent and T-independent antigens, Factors responsible for immunogenicity, Adjuvants, Super-antigens, Structure and function of antibody, Antibody classes, VDJ rearrangements, Monoclonal and chimeric antibodies, Major Histocompatibility: Structure and Functions of MHC I & II molecules, Antigen processing and presentation, Inflammatory response, Complement System	
Unit III: Generation of Immune Response and Vaccines	18 hrs
Primary and Secondary Immune Response, Generation of Humoral Immune Response (Plasma and Memory cells), Generation of Cell Mediated Immune Response (Self MHC restriction, T cell activation, Co- stimulatory signals), Killing Mechanisms by CTL and NK cells, Types of autoimmunity and hypersensitivity with examples, Immunodeficiencies - Animal models (Nude and SCID mice), SCID, DiGeorge syndrome, Chediak- Higashi syndrome, tumor antigens, Vaccines: Active and passive immunization, Vaccine types (Live but attenuated, Killed, Subunit, Recombinant, DNA and Peptide)	
Unit IV: Immunological Techniques	18 hrs

Principles of Precipitation, Agglutination, Immunodiffusion, Immunoelectrophoresis, Immunoassays, ELISA, ELISPOT, Western blotting, Immunofluorescence, Flow cytometry, fluorescence activated cell sorting analysis, microarrays to assess gene expression	
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List of Experiments:

1. Identification of human blood groups.
2. Total Leukocyte Count of the given blood sample.
3. Differential Leukocyte Count of the given blood sample.
4. Separation of serum from the given blood sample.
5. Immunodiffusion by Ouchterlony method.
6. DOT ELISA.
7. Immunoelectrophoresis.

Course Learning Outcomes:

1. Students will be able to explain the role of immune cells and their role in body defense mechanism
2. Students will be able to devise strategies to combat infection or diseases produced by altered self.
3. Students will develop ability to use this knowledge in the processes of immunization, antibody engineering, vaccine development, transplantation, and diseases.
4. Students will be able to demonstrate immunological techniques

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
J. Owen, J. Punt, S. Stranford	Kuby Immunology (8 th Edition)	WH Freeman and Company, USA	2012		
D. Male, J. Brostoff, D. Roth, I. Roitt	Immunology (8 th Edition)	Saunders, Elsevier, USA	2012		
K. Murphy	Janeway's Immunobiology (8 th Edition)	Garland Science, USA	2011		

A. Abbas, A. Lichtman, S. Pillai	Cellular and Molecular Immunology (8 th Edition)	Saunders, Elsevier, USA	2014		
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BTY206: Bioprocess Technology

L	T	P	Total Credits
4	0	2	6

Course Objectives: Study bioprocess technology to comprehend the principles and applications of biological systems in industrial processes, enabling the optimization of bioproduction methods for various products.

Course content and syllabus

	Teaching Hours
Unit I: Introduction to Bioprocess Technology and Media Design for Fermentation	18 hrs
Bioprocess vs chemical processing, advantages, disadvantages, Substrates for bioconversion process, Isolation, Preservation Techniques and Maintenance methods of Industrial Microorganisms, Cell culture technique, Media composition and design, Media type, Commercial media, Inoculum development and aseptic transfers, Criteria for inoculums transfer,(media formulation RSM, CCD,)	
Unit II: Process Technology for Production of Primary Metabolites	18 hrs
Ethanol: production by batch and continuous process by various technologies. Determination of fermentation efficiency, distillation efficiency and overall efficiency of ethanol production, recovery, uses, glucose effect etc. Power alcohol – definition, uses, merits and demerits of various technologies for its production. Amino Acid: Lysine: Indirect and direct fermentation Biomass: Bakers and distillers yeast production using various raw materials, “bios” factors for growth, Crabtree effect, harvesting, different forms and uses.	
Unit III: Process Technology for Production of Secondary Metabolites	18 hrs

Production of secondary metabolites – penicillin, streptomycin, Penicillin: Classification, various penicillin as precursor and 'R' – side chain, penicillinase, 6-APA, penicillin production, harvest and recovery, uses of various forms etc. Streptomycin: chemical structure, production, harvest and recovery, use by-product of streptomycin fermentation etc.	
Unit IV: Microbial production of industrial enzymes	18 hrs
Glucose isomerase, Amylase, Lipase, Protease	

List of Experiments -with basic instructions (Total Teaching = 72 hrs)

1. Isolation of industrially important bacteria for microbial processes.
2. Estimation of Reducing sugar by DNS method of unknown sample and establish calibration curve.
3. Estimation of ethanol by potassium dichromate methods and establish calibration curve.
4. Establish growth profile of bacteria with respect of optical density and maximum specific growth rate of bacteria
5. Determination of Thermal Death Point and Thermal death time of bacteria for design of a sterilizer.
6. Cultivation of microorganism in batch process
7. Ethanol production using grape juice
8. Demonstration of bioreactor

Course Learning Outcomes:

1. Design and formulation of production media for fermentation,
2. Upstream and downstream techniques of primary and secondary product
3. Production technology of biomass, primary and secondary metabolites

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
M.L. Shuler and Fikret. Kargi, 2nd edition	Bioprocess Engineering	Pearson Education Limited.	2013	9780136060659	957

	Basic Concepts,				
W. Crueger and A. Crueger,	Biotechnology : A Textbook of Industrial Microbiology,	Sinauer Associates	1990	0878931317	541

MBO203: Microbial Physiology and Metabolism

L	T	P	Total Credits
4	0	2	6

Course content and syllabus

Course Objectives: Teach students basics of microbial genetics and impart knowledge about the mechanism of genetic exchange and transposable elements

	Teaching Hours
Unit I: Microbial growth and effect of environmental factors on growth	18 hrs
<p>Definitions of growth, measurement of microbial growth, Batch culture, Continuous culture, generation time and specific growth rate, synchronous growth, diauxic growth curve</p> <p>Microbial growth in response to environment -Temperature (psychrophiles, mesophiles, thermophiles, extremophiles, thermodurics, psychrotrophs), pH (acidophiles, alkaliphiles), solute and water activity (halophiles, xerophiles, osmophilic), Oxygen (aerobic, anaerobic, microaerophilic, facultative aerobe, facultative anaerobe), barophilic. Microbial growth in response to nutrition and energy – Autotroph/Phototroph, heterotrophy, Chemolithoautotroph, Chemolithoheterotroph, Chemoheterotroph, Chemolithotroph, photolithoautotroph, Photoorganoheterotroph.</p> <p>Passive and facilitated diffusion Primary and secondary active transport, concept of uniport, symport and antiport</p>	
Unit II: Chemoheterotrophic Metabolism	18 hrs
<p>Aerobic Respiration: Concept of aerobic respiration, anaerobic respiration and fermentation. Sugar degradation pathways i.e. EMP, ED, Pentose phosphate pathway TCA cycle, Electron transport chain: components of respiratory chain, comparison of mitochondrial and bacterial ETC, electron transport phosphorylation, uncouplers and inhibitors.</p> <p>Anaerobic respiration and fermentation: Anaerobic respiration with special reference to dissimilatory nitrate reduction (Denitrification; nitrate /nitrite and nitrate/ammonia respiration; fermentative nitrate reduction)</p>	

Fermentation - Alcohol fermentation and Pasteur effect; Lactate fermentation (homofermentative and heterofermentative pathways), concept of linear and branched fermentation pathways.	
Unit III: Chemolithotrophic and Phototrophic Metabolism	18 hrs
Introduction to aerobic and anaerobic chemolithotrophy with an example each. Hydrogenoxidation (definition and reaction) and methanogenesis (definition and reaction) Introduction to phototrophic metabolism - groups of phototrophic microorganisms, anoxygenic vs. oxygenic photosynthesis with reference to photosynthesis in green bacteria, purple bacteria and cyanobacteria	
Unit IV: Nitrogen Metabolism	18 hrs
Introduction to biological nitrogen fixation Ammonia assimilation Assimilatory nitrate reduction, dissimilatory nitrate reduction, denitrification	

List of Experiments -with basic instructions (Total Teaching = 72 hrs)

1. Study and plot the growth curve of *E. coli* by turbidometric and standard plate count methods.
2. Calculations of generation time and specific growth rate of bacteria from the graph plotted with the given data
3. Effect of temperature on growth of *E. coli*
4. Effect of pH on growth of *E. coli*
5. Effect of carbon and nitrogen sources on growth of *E. coli*
6. Effect of salt on growth of *E. coli*
7. Demonstration of alcoholic fermentation
8. Demonstration of the thermal death time and decimal reduction time of *E. coli*.

Course Learning Outcomes:

1. Study the types of microbial growth and effect of environmental factors.
2. Understand the mechanism of nutrients uptake and transport in micro-organisms.
3. Study different types of growth in micro-organisms in response to nutrition and energy.
4. Classify the microbes on the basis of metabolic processes and their energy requirements.
5. Perceive knowledge of microbial nitrogen metabolism.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Madigan MT, and Martinko JM (2014)	Brock Biology of Microorganisms. 14th edition	Prentice Hall International Inc.	2014	9781292018317	1030
Moat AG and Foster JW	Microbial Physiology 4 th Edition	John Wiley & Sons	2002	0471394831 978-0471394839	736
Wiley JM, Sherwood LM, and Woolverton CJ	Prescott's Microbiology . 9 th edition.	McGraw Hill Higher Education.	2013	9780073402406 0073402400	2272
Pelczar Jr MJ, Chan ECS, and Krieg NR.	Microbiology. 5th edition	Tata McGraw Hill.	1993	0070492581, 9780070492585	957

BTY207: RECOMBINANT DNA TECHNOLOGY

L	T	P	Total Credits
3	0	1	4

Course content and syllabus

Course Objective: To teach methods of DNA manipulations, cloning and gene editing

	Teaching Hours
Unit I: Gene Cloning and DNA Analysis	18 hrs
Polymerase chain reaction, DNA modifying enzymes: polymerases, kinases, ligases, phosphatases; Primers designing, Purification of DNA fragments, Restriction enzymes, DNA ligation, Vectors, DNA Transformation, GENOMIC DNA and Plasmid Isolation, Restriction digestion and DNA Analysis by gel electrophoresis.	
Unit II: Vectors for Gene Cloning and DNA Manipulation	9 hrs
Cloning vectors based on E. coli plasmids, Plasmid copy number control, Cloning vectors based on M13 bacteriophage, Cloning vectors based on λ bacteriophage, λ and other high-capacity vectors enable genomic libraries to be constructed, Vectors for other bacteria, Bacterial Artificial chromosomes (BACs); Vectors for yeast and other fungi, Yeast artificial chromosomes (YACs), Cloning vectors for higher plants, Tobacco Mosaic Virus (TMV); Cloning vectors for animals. Problem of Plasmid incompatibility, The problem of selection, Direct selection, Identification of a clone from a gene library, Methods for clone identification.	
Unit III: Cloning a Specific Gene	18 hrs
Transduction, conjugation and transfection, Types of plasmids, Recombinant Bacterial strains for bioremediation; online servers/software for DNA and protein analysis: Acquiring DNA sequence encoding the protein of interest (for example GFP) from online database like PUBMED and PDB. Analysis of DNA sequence for presence of internal restriction digestion sites etc.	
Unit IV: Advanced Cloning Techniques	9 hrs
Homologous recombination, Molecular mechanism of RecBCD, RecA, RuvA-B, Holliday Model; Non-homologous End Joining (NHEJ) versus Homologous DNA recombination; Positive and negative selection, Zinc finger nucleases (ZFN), Transcription activator-like effector nucleases (TALENs), Discovery of adaptive immunity, The CRISPR-Cas9 (clustered regularly interspaced short palindromic	

repeats) system, Methods to create gene-knock out animal model systems. Cre-LoxP recombination system	
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List of Experiments

1. Acquiring DNA sequence encoding the protein of interest (for example GFP) from online database like Genbank and Uniprot. Analysis of DNA sequence for presence of internal restriction digestion sites etc using softwares like gene runner.
2. Primer designing: Designing of 5' forward and 3' reverse complementary primers containing appropriate restriction digestion sites, affinity tags (penta-His etc.).
3. PCR amplification of the DNA segment of interest from a suitable source, purification of the PCR product.
4. Restriction digestion, and subsequent ligation into the suitable bacterial expression vector (also containing an antibiotic resistant marker) of interest.
5. Preparation of competent cells and transformation into suitable competent cells (BL21 etc.).
6. Selection of the antibiotic resistant single colony.
7. Plasmid isolation from the transformed cells and sequencing it to confirm the sequence of cloned DNA segment of interest.

Course Learning Outcomes:

Students will be able to:

1. Understand basic concepts of DNA manipulation.
2. Understand the procedure of gene cloning
3. Have a thorough understanding of vectors
4. Perceive knowledge of advanced gene editing methods

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
J. Sambrook, E. F. Fritsch, and T. Maniatis, 2nd Edn.,	Molecular cloning: a laboratory manual,	Cold Spring Harbor Laboratory Press	1989		
T.A. Brown	Gene Cloning and DNA Analysis - An	Wiley - Blackwell	2010	9781405181730	338

	introduction				
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PHY213: Fundamentals of Physics

L	T	P	Total Credits
2	0	0	2

Course Objectives: Aim of this course is to introduce the students about fundamentals of graduate level Physics, which forms the basis of all Applied Science specifically physical optics, mechanics, dynamics, and acoustics relating human body

Course content and syllabus

Unit I: Interference, diffraction, and polarization (10)	HRS
Young's double slit experiment, Huygen's principle, Superposition principle, Analysis of interference (constructive and destructive) and conditions for sustained interference, Interference in thin parallel and wedge-shaped films, Newton's rings, introduction to diffraction: Fresnel and Fraunhofer diffraction, Rayleigh criterion and Resolving power and dispersive power of grating, Polarization of Light, Law of Malus, Brewster's Law, Circularly and Elliptically Polarized Light, Half and Quarter Wave Plates	9 hrs
Unit II: Lasers (8)	
Introduction of Lasers, Induced Absorption, Spontaneous and Stimulated Emission, Einstein Coefficients, Population inversion, Fundamental of Lasers, Types of Pumping, Concept of Three and Four Level Lasers, Construction and Working Lasers, Properties of Laser and its applications	9 hrs
Unit III: Mechanics and dynamics of a human body (12)	
Mechanics: Muscular Action, Friction, Energetics, Model of Walking, Material Components of the Body, Bone, Ligaments and Tendons, Cartilage, Elastic Properties, Basic Stress-Strain Relationships, Fluid mechanics: Characteristic Pressures in The Body, Physics of Pressure and Flow of Fluids, Law of Laplace, Fluids in Motion, Equation of Continuity, Bernoulli's Equation, Viscous Flow and Poiseuille's Law, Thermodynamics: First and second laws of thermodynamics, Concept of free energy, entropy, heat content of food, bomb calorimetry	9 hrs
Unit IV: Waves and human body (6)	

Speed and Properties of Sound Waves, Intensity of Sound Waves, Sound propagation from one Medium to Another, Speech Production, Types of Sounds, Hearing, Other	9 hrs
Vibrations of the Body, Cardiac and Other Sources of Sounds	

Course Learning Outcomes:

1. Understand the fundamental principles underlying phenomena of interference, diffraction, and polarization
2. Understanding on the properties, construction, and applications of laser
3. Understand and analyse the mechanical and dynamical aspects of the different components of a human body
4. Understand and analyzing basics of sound with its impact on the body

Text/Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
Irving P. Herman	Physics of the Human Body	Springer , ISSN1618-721	2006	978-3540817062
W. Hughes B	Aspects of Biophysics	John Willey and Sons	1979	978-0471019909
R.K. Hobbie	Intermediate Physics in Biology and Medicine	Springer	2001	978-3319126814
Halliday, Resnick and Walker	Fundamentals of Physics	Wiley India Pvt Ltd	2006	978-8126514427
Brijlal, Subramanyam & N Subrahmanyam	Principle of Optics	S. Chand publishing, 25th edition, 2012	2006	978-8121926119

Ghatak, Ajay	Optics	Tata McGraw-Hill	4th Edition	978933922090 7
Jenkins F A, White H E	Fundamentals of optics	Mcgraw hill	4th Edition	978007256191 3

Programme structure for B.Sc. (H) Microbiology- 3 years (5th Semester)

Sr. No	Course Code	Course Title	Course Type	Credits					Credit Units
				L	T	P	FW	SW	
1	HGM301	Molecular Biology	Core Course	4	0	2	0	0	6
2	MBO301	Food and Dairy Technology	Core Course	4	0	2	0	0	6
3.	MBO302	<u>Students will choose any two of the given choices*</u>	Specialization Elective Course	4	0	0	0	0	4
		1. Microbial Genetics		4	0	0	0	0	4
4.	MBO303	2. Nanomedicine							
		3. Virology							
		4. Archaea and Extremophiles							
		5. MOOC							
5.		<u>Students will choose any two of the given choices**</u>	Skill Enhancement Course						
		1. Cell Signalling							
		2. Research Methodology							
		3. Biowarfare and Bioterrorism	2	0	0	0	0	2	
		4. Programming in Python	2	0	0	0	0	2	
		5. MOOC							

Total Credits

24

*The Specialization Elective Courses of 5th and 6th Semesters will be pooled together.

**The Skill Enhancement Courses of 5th and 6th semesters will be pooled together.

The Specialization Elective Course and the Skill Enhancement Course can also be taken through MOOC. A maximum of 4 credits per semester can be taken through MOOC.

HGM301: Molecular Biology

L	T	P	Total Credits
4	0	2	6

Course content and syllabus

Course Objectives: To teach the fundamentals of DNA replication, transcription and translation

	Teaching Hours
Unit I: Genes and Genomes	15 hrs
<p>The History and Birth of Molecular Biology. Relationships between genotype and phenotype. Contributions of Nobel Laureates in the area of Molecular Biology</p> <p>Genes and Genomes: Molecular definition of gene. Organization of genes on chromosomes. Repetitive DNA. Simple sequence DNA. Interspersed-Repeat DNA and mobile DNA elements.</p> <p>Chromosome structure: Bacterial chromatin and specific proteins to condense bacterial DNA.</p> <p>Nucleosomes. Chromatin organization in eukaryotes. Functional Rearrangements in chromosomal DNA. Extra-nuclear genomes, Specific notations, conventions and terminologies used in genetics</p>	
Unit II: DNA Replication, Damage and Repair	21 hrs
<p>DNA replication is semi-conservative and bi-directional.</p> <p>DNA replication in bacteria: Initiation, elongation and termination of bacterial DNA replication. Enzymes involved in DNA replication.</p> <p>Eukaryotic DNA replication machinery. Initiation, elongation and termination of replication. Telomeres and Telomerase. Leading strand problem in replication.</p> <p>DNA replication in Archaea</p>	

DNA damage and repair mechanisms	
Unit III: Transcription	18 hrs
<p>RNA Transcription in bacteria and eukaryotes</p> <p>RNA and Transcription: Types of RNA. Types of RNA polymerase and structure; Molecular apparatus and events during prokaryotic and eukaryotic RNA synthesis. Post—transcriptional modifications of transcripts. Processing of different types of RNA. RNA editing. Formation of spliceosome complex. Inhibitors of RNA metabolism and their mechanism of action; RNA degradation.</p>	
Unit IV: Protein Translation	18 hrs
<p>Genetic code: Its deciphering, degeneracy and general features.</p> <p>tRNA, aminoacylation of tRNA, tRNA identity and aminoacyl tRNA synthetases. Structure of ribosomes, and its assembly and disassembly. Codon: anti-codon base pairing, Wobble hypothesis</p> <p>Translation in Prokaryotes: formation of initiation complex, initiation factors, elongation, elongation factors, and termination.</p> <p>Translation in Eukaryotes: formation of initiation complex, initiation factors, elongation, elongation factors and termination.</p> <p>Translation proof-reading, translation inhibitors.</p> <p>Post-translation modifications of proteins and their effect on their structure and function.</p> <p>Protein targeting: Signal sequence and targeting of proteins to specific cellular locations.</p>	

List of Experiments -with basic instructions (Total Teaching = 72 hrs)

1. Verification of Chargaff's rule by paper chromatography.
2. Ultraviolet absorption spectrum of DNA and RNA.
3. Determination of DNA and RNA concentration by A₂₆₀nm.
4. Determination of the melting temperature and GC content of DNA.
5. To study the viscosity of DNA solutions.
6. Isolation of chromosomal DNA from E. coli/plant/yeast/animal cells.
7. Recombinant Protein Expression and Purification

Course Learning Outcomes: at the end of this course, students will learn about:

1. History and development of molecular biology, structure of genome and terminologies used in molecular genetics
2. DNA replication in bacteria, archaea and eukaryotes
3. Mechanism of transcription in bacteria and eukaryotes.
4. Mechanisms of translation and bacteria and eukaryotes.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Watson, J.D., Baker, T.A., Stephen, P.B., Alexander, G., Levine, M., Losick R.	Molecular Biology of the Gene	Pearson Education	7 th Ed	978- 9332585478	912
Tropp, B.E.	Molecular Biology Genes to Proteins	Jones and Bartlett	4 th Ed	978-93-80853- 49-9	1096
Lewin, B.	Genes XI	Jones and Bartlett	10 th ED/2014	97814496590 59	940

MBO301: Food and Dairy Technology

L	T	P	Total Credits
4	0	2	6

Course Objectives: Teach basic principles and methods of food processing, preservation and process technology of milk and milk products and will also obtain knowledge about laws and standards for milk and milk products.

Course content and syllabus

	Teaching Hours
Unit I: Basic principles and methods of food processing and preservation.	13 hrs
Emerging Technologies in food processing. Food additives and preservatives. Food laws and standards. Effect of processing on acceptability and nutritive value of food.	
Unit II: Physico-chemical properties and structure of milk and milk constituents.	13 hrs
Chemical and microbial spoilage of milk and milk products; Fluid milk Processing, packaging and distribution. Common dairy processes – cream separation (standardization), pasteurization, sterilization and Homogenization.	
Unit III: process technology for various milk products	18 hrs
Process technology for manufacture of evaporated milk, condensed milk, dried milk, malted milk, infant and baby foods, ice-cream, cheese, butter, fermented milk and indigenous dairy products. Fermentation of foods	

Unit IV: Laws and standards for milk and milk products.	28 hrs
Laws and standards for milk and milk products. Technological processes for industrially manufactured foods of commercial importance, from plant and animal origin. Gas packaging and modified atmosphere Package design. Shelf life prediction of foods in packages. Quality control in Food packaging. Product safety and packaging regulations. Government schemes and incentives for promotion of entrepreneurship. Government policy on Small and Medium Enterprises (SMEs) / SSIs. Export and Import Policies relevant to horticulture sector. Venture capital. Contract farming and joint ventures, public-private partnerships. Overview of horticulture industry. Characteristics of Indian horticultural processing and export industry. Social Responsibility of Business.	

List of Experiments -with basic instructions (Total Teaching = 60 hrs)

1. Basic labs on sterilization, food pasteurization and homogenization
2. Preservation of a microbial strain.
3. Precipitation of protein (casein) from milk with an acid.
4. Determination of Milk Fat by Gerber Method.
5. Determination of Specific Gravity of Milk.
6. To perform Microbial Analysis of Foods.
7. Determination of Milk Fat (by Acid Digestion Method) in Cheese.
8. DETERMINATION OF LACTOSE BY COLORIMETRIC METHOD IN MILK BASED SWEETS.

Course Learning Outcomes: impart knowledge and technical proficiency in:

1. Clean milk production, handling and processing of milk
2. Manufacture of western and indigenous dairy products
3. Testing and quality control of milk and milk products
4. Marketing and economical aspects
5. Managing small and medium enterprises

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Pelczar, M.J. Jr., Chan ECSand Krieg, N.R.	Microbiology: Concepts and Applications	New York; Madrid : McGraw- Hill,	1993	0070492581, 97800704925 85	957

Cappucino, J.G.	Microbiology- A laboratory manual	Hoboken, N.J.: Pearson	2020	0135188997, 97801352039 96,	541
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MBO302: Microbial Genetics

L	T	P	Total Credits
4	0	0	4

Course Objectives: Teach students basics of microbial genetics and impart knowledge about the mechanism of genetic exchange and transposable elements

Course content and syllabus

	Teaching Hours
Unit I: Genome Organization and Mutations	18 hrs
<p>Genome organization: <i>E. coli</i>, <i>Saccharomyces</i>, <i>Tetrahymena</i>. Organelle genome: Chloroplast and Mitochondria.</p> <p>Mutations and mutagenesis: Definition and types of Mutations; Physical and chemical mutagens; Molecular basis of mutations; Functional mutants (loss and gain of function mutants); Uses of mutations.</p> <p>Reversion and suppression: True revertants; Intra- and inter-genic suppression; Ames test; Mutator genes.</p>	
Unit II: Plasmids	18 hrs
<p>Types of plasmids – F plasmid, R Plasmids, colicinogenic plasmids, Ti plasmids, linear plasmids, yeast- 2 μ plasmid, Plasmid replication and partitioning, Host range, plasmid-incompatibility, plasmid amplification, Regulation of copy number, curing of plasmids</p> <p>Phage Genetics: Features of T4 genetics , Genetic basis of lytic versus lysogenic switch of phage lambda</p>	
Unit III: Mechanisms of Genetics Exchange	18 hrs
<p>Transformation - Discovery, mechanism of natural competence</p> <p>Conjugation - Discovery, mechanism, Hfr and F' strains, Interrupted mating technique and time of entry mapping</p>	
<p>Transduction - Generalized transduction, specialized transduction, LFT & HFT lysates, Mapping by recombination and co-transduction of markers</p>	

Unit IV: Transposable Elements	18 hrs
Prokaryotic transposable elements – Insertion Sequences, composite and non-composite transposons, Replicative and Non replicative transposition, Mu transposon Eukaryotic transposable elements - Yeast (Ty retrotransposon), Drosophila (P elements), Maize (Ac/Ds) Uses of transposons and transposition	

Course Learning Outcomes: at the end of the students will learn about

1. Basics of microbial genetics including organization of genome of various microorganisms
2. Mutations and their importance
3. Roles and significance of transposable elements
4. Mechanisms of genetic exchange

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Snyder, L., Peters, J.E., Henkin, T.M., and Champness, W.	Molecular Genetics of Bacteria	ASM Press	4 th Ed	978-1-55581- 892-0	707
Klug WS, Cummings MR, Spencer, C, Palladino, M.	Concepts of Genetics	Pearsons	11 th Ed.	978- 9353940409	896
Pierce BA	Genetics: A Conceptual Approach	WH Freeman	7 th Ed.	978- 1319308315	976

Nanomedicine

L	T	P	Total Credits
4	0	0	4

Course Objective: To make students acquainted with the fundamental concepts of nanotechnology and develop an understanding to employ its principles in biomedical applications.

Course content and syllabus

	Teaching Hours
Unit I: Introduction to nanomaterials	18 hrs
Importance of "Nano" dimension, size matters: bulk vs nanomaterials, nanotechnology exists in nature, brief history of nanotechnology, concept of dimensionality of nanomaterials, effect of 'nano' scale on material properties (electrical, thermal, mechanical, optical, chemical), quantum structures, quantum confinement, classification of nanostructured materials, surface effects of nanomaterials, nanocomposites	
Unit II: Synthesis and Characterization of Nanomaterials	18 hrs
Bottom-up and top-down approaches, physical and chemical methods: mechanical milling, laser ablation, arc discharge, chemical vapor deposition, physical vapor deposition, wet chemical synthesis of nanoparticles, self-assembly, biological synthesis of nanomaterials	
Unit III: Bionanotechnology	18 hrs
Surface functionalization of nanomaterials for biological applications, nano-antimicrobials, viral nanotechnology, Biological nanomachines: protein and DNA, peptide nanotechnology, DNA nanotechnology, cellular uptake mechanisms of nanomaterials	
Unit IV: Nanomaterials Applications in Biology and Nanotoxicity	18 hrs
Polymeric biomaterials, lipid nanoparticles for drug delivery applications, nanoparticles for bioimaging, cancer therapeutics, and tissue engineering applications, stimuli-responsive nanoparticles, nano-artificial cells, nanomaterials for organ printing, nanotoxicology	

Course Learning Outcomes: Students will be able to

1. Comprehend the concept of "nanotechnology" and its interdisciplinary aspects.

2. Learn various approaches of synthesizing nanomaterials, their advantages, and limitations.
3. Gain knowledge about various techniques used for characterizing nanomaterials.
4. Comprehend the importance of engineered nanomaterials for biomedical, and therapeutic applications.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
G. Cao and YngWang	Nanostructures and Nanomaterials: Synthesis, Properties and Applications	World Scientific, 2011	2004	9814322504, 9789814322508	581
C. M. Niemeyer, C. A. Mirkin	Nanobiotechnology; Concepts, Applications and Perspectives	Wiley-VCH	1 st ED/2004	352730658, 978- 352730658	491
B. S. Murthy, P. Shankar, B. Raj, B. B. Rath and J. Murday	Textbook of Nanoscience and Nanotechnology	Universities Press-IIM	2012	9788173717383, 3, 978- 8173717383	256
T. Pradeep	Nano:The Essentials	Tata McGraw-Hill Publishing Company Ltd.	2007	0070617880, 9780070617889	461

MBO303: Virology

L	T	P	Total Credits
4	0	0	0

Course Objectives: This course will give an overview of important viruses, their mode of replication and methods to control them.

Course content and syllabus

	Teaching Hours
Unit I: Nature and Properties of Viruses	18 hrs
Introduction: Discovery of viruses, nature and definition of viruses, general properties, concept of viroids, virusoids, satellite viruses and Prions. Theories of viral origin. Structure of Viruses: Capsid symmetry, enveloped and non-enveloped viruses. Isolation, purification and cultivation of viruses. Viral taxonomy: Classification and nomenclature of different groups of viruses.	
Unit II: Bacteriophages	18 hrs
Diversity, classification, one step multiplication curve, lytic and lysogenic phages (lambda phage) concept of early and late proteins, regulation of transcription in lambda phage.	
Unit III: Viral Replication and Transmission	18 hrs
Modes of viral transmission: Persistent, non-persistent, vertical and horizontal. Salient features of viral Nucleic acid : Unusual bases (TMV, T4 phage), overlapping genes (ϕ X174, Hepatitis B virus), alternate splicing (HIV), terminal redundancy (T4 phage), terminal cohesive ends (lambda phage), partial double stranded genomes (Hepatitis B), long terminal repeats (retrovirus), segmented (Influenza virus), and non-segmented genomes (picornavirus), capping and tailing (TMV). Viral multiplication and replication strategies: Interaction of viruses with cellular receptors and entry of viruses. Replication strategies of viruses as per Baltimore classification (ϕ X 174, Retroviridae, Vaccinia, Picorna), Assembly with example of Polio virus and T4 phage, maturation and release of virions.	
Unit IV: Viral Diseases, prevention and application of viruses	18 hrs

<p>Introduction to oncogenic viruses: Types of oncogenic DNA and RNA viruses: Concepts of oncogenes and proto-oncogenes.</p> <p>Antiviral compounds and their mode of action. Interferon and their mode of action.</p> <p>General principles of viral vaccination.</p> <p>Application of virology: Use of viral vectors in cloning and expression, Gene therapy, Phage display and phage therapy.</p>	
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Course Learning Outcomes: at the end of the course the students will:

1. Get an exhaustive account of viruses, their structure and classification
2. Learn about replication of viruses
3. Viral diseases and methods to control them
4. Various applications of virology

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Dimmock, NJ, Easton, AL, Leppard, KN	Introduction to Modern Virology	Blackwell Publishing Ltd	6 th Ed	978-1405171120	536
Flint SJ, Enquist, LW, Krug, RM, Racaniello, VR, Skalka, AM	Principles of Virology, Molecular biology,	ASM Press	2 nd Ed (2000)	978-1555811273	820
	Pathogenesis and Control				
Carter J and Saunders V	Virology: Principles and Applications	Wiley	2 nd (2013)	978-1119991427	394

Archaea and Extremophiles

L	T	P	Total Credits
4	0	0	0

Course Objectives: Investigate archaea and extremophiles to understand their unique biology, ecological roles, and potential applications in biotechnology.

Course content and syllabus

	Teaching Hours
Unit I: Basics on extremophiles	18 hrs
Concept of extremophiles v/s conventional microbial forms & archaea, habitats in universe, eco- niches, communities and community associations, biofilms, microbial community analysis of extreme environments using various molecular approaches (DGGE, cloning and next generation sequencing, functional genomics and transcriptomics).	
Unit II:	18 hrs
Occurrence, Physiological features, adaptation strategies of various extremophilic microbes: a) anearobes, barophiles/ peizophiles, cryophiles & thermophiles; b) oligotrophs, osmophiles, halophiles & xerophiles; [Text Wrapping Break]c) radiophiles, metallophiles & xenobiotic utilizers; d) alkaliphiles/ basophiles, acidophiles. Potential applications of extremophilic microbes.	
Unit III: Microbes in toxic environments	18 hrs
Microbes in toxic environments: acid mine drainage, waste containing cyanides, xenobiotics, pesticides, heavy metals and radio isotopic materials,	
Unit IV: Applications and case studies	18 hrs
extremozymes and their applications, field and case studies.	

Course Learning Outcomes:

1. Know the types of microbial diversity flourish in extreme environments.
2. Understand how organisms cope under extreme living conditions with biochemical and molecular adaptation of extremophilic microorganisms.
3. Understand modern techniques used for exploration of unculturable extremophiles
4. Understand potential application of extremozymes in various industries and in functional genomics.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Brock, T. D.	Thermophilic Microorganisms and Life at High Temperatures	Springer, New York.	1978	978-1-4612- 6284-8	468
Fred A Rainey and Aharon Oren	Extremophiles	Academic press	2006	978012521536 7	838

Cell Signalling

L	T	P	Total Credits
2	0	0	2

Course Objectives: Explore cell signaling to understand molecular mechanisms governing cellular communication, facilitating insights into physiological processes, disease pathways, and therapeutic targets.

Course content and syllabus

	Teaching Hours
Unit I: Introduction to Cell Signaling	9 hrs
Modes of signaling, signaling molecules, Intracellular and cell surface receptors, Hormonesignaling	
Unit II: GPCR & RTK	9 hrs
G-protein mediated signaling, second messengers, receptors tyrosine kinases, Ras-MAPKpathway, JAK-STAT pathway, PI3K-AKT pathway,	
Unit III: Signaling through other pathways	9 hrs
Integrins,cadherins,Hedgehog,Notch,Heat shock and ER stress response, Serine/Threoninepathways	
Unit IV: Aberrant signaling	9 hrs
Cancer, Notch signaling dependent Diseases, Hedgehog signaling dependent Diseases,Diabetes	

Course Learning Outcomes:

1. Differentiate structure, receptors, and mechanism of actions of hormones.
2. Describe pathways of cellular signaling, cross-talk and regulation.
3. Discuss how disruptions in cellular signaling may lead to disease, and illustrate with selected examples.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Bruce, Alberts and Alexander Johnson and Julian Lewis, and MartinRaff	Molecular biology of the cell	Garland Science;	6th	978- 0815344322	1342
Rakesh Srivastava	Apoptosis, cell signalling and human diseases	Humana Press	1st	978158829882 9	395
Berg J.M., Tymoczko J.L., Stryer L.	Biochemistry	WH Freeman &Company	5th	13: 978-1- 4641- 2610-9	1023

Research Methodology

L	T	P	Total Credits
2	0	0	2

Course Objectives: Examine research methodology to develop critical skills in designing, executing, and analyzing research studies across various disciplines, fostering a comprehensive understanding of scientific inquiry and advancing proficiency in evidence-based research practices.

Course content and syllabus

	Teaching Hours
Unit I: Basic Concepts	9 hrs
Research process, problem identification, research designs, informal experimental designs. Completing randomized design, randomized block design, latin square design, factorial designs	
Unit II: Sample collection	9 hrs
Random sampling, complex random sampling, non-probability sampling, measurement and scaling techniques. Data collection.	
Unit III: Research Presentation	9 hrs
The students will be taught to present their work in written form and also how to make effective power point presentation	
Unit IV: Literature Survey	9 hrs
The students will be required to review literature in their respective disciplines and submit an assignment for evaluation.	

Course Learning Outcomes:

1. Teach students importance of research conceptualization and planning

2. Teach student how to make effective written and spoken presentations
3. Teach students how to read a research paper

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Kothari, C.R	Research Methodology: Methods and Techniques	New Age International Publishers	4 th Ed.	978- 9386649225	480
Arya., P.P. and Pal, Y	Research Methodology in Management: Theory and Case Studies	Deep and Deep Publishers	2011	978- 8184503715	

Biowarfare and Bioterrorism

L	T	P	Total Credits
2	0	0	2

Course Objectives: Analyze biowarfare and bioterrorism to understand the threats posed by biological agents, explore strategies for detection, prevention, and response, and preparedness against bioterrorist threats.

Course content and syllabus

	Teaching Hours
Unit I: Introduction	9 hrs
History of Biowarfare. Difference between biowarfare and bioterrorism. Laws preventing the use of Bioweapons	
Unit II: Agents of Biowarfare and Bioterrorism	9 hrs
Various biological agents (bacteria and viruses) that can be used as bioweapons, their properties, mode of spread, infection, incubation period, symptoms, and current treatment strategies.	
Unit III: Dissemination and Detection of Biological Agents	9 hrs
Modes of dissemination or delivery of biological Agents: by air through aerosol spray; through explosives (missile, bombs, artillery, etc), contamination of food and water; injected or absorbed through the skin Methods to detect and identify biological agents.	
Unit IV: Mitigation Strategies	9 hrs
Public Health and emergency response preparedness. Role of antimicrobials, vaccines, antibodies, immune modulators, and other medications in mitigation. Uses of different biomaterials as a protective cover.	

Course Learning Outcomes:

1. Will learn about the history of biological warfare
2. Understand the agents used as biological weapons
3. Delivery of Biological Agents
4. Methods to detect and identify biological agents.
5. Public Health strategies to mitigate effects of biological weapons

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Dando, M.R.	Bioterror and Biowarfare - A Beginner's Guide	OneWorld Publications	2006	978- 1851684472	256
Boyle, F.	Biowarfare and Terrorism	Clarity Press	2005	978- 0932863461	139

Programming in Python Lab

L	T	P/S	SW/FW	TOTAL CREDIT UNITS
0	0	4	0	2

Course Contents/syllabus:

List of Experiments (Total: 72 Hours)

1. Compute sum, subtraction, multiplication, division and exponent of given variables input by the user.
2. Compute area of following shapes: circle, rectangle, triangle, square, trapezoid and parallelogram.
3. Compute volume of following 3D shapes: cube, cylinder, cone and sphere.
4. Compute and print roots of quadratic equation $ax^2+bx+c=0$, where the values of a, b, and c are input by the user.
5. Print numbers up to N which are not divisible by 3, 6, 9,, e.g., 1, 2, 4, 5, 7,....
6. Write a program to determine whether a triangle is isosceles or not?
7. Print multiplication table of a number input by the user.
8. Compute sum of natural numbers from one to n number.
9. Print Fibonacci series up to n numbers e.g. 0 1 1 2 3 5 8 13.....n
10. Compute factorial of a given number.
11. Count occurrence of a digit 5 in a given integer number input by the user.
12. Print Geometric and Harmonic means of a series input by the user.
13. Evaluate the Arithmetic expressions.
14. Print all possible combinations of 4, 5, and 6.
15. Determine prime numbers within a specific range.
16. Count number of persons of age above 60 and below 90.
17. Compute transpose of a matrix.
18. Perform following operations on two matrices.
 - 1) Addition 2) Subtraction 3) Multiplication
19. Count occurrence of vowels.
20. Count total number of vowels in a word.
21. Determine whether a string is palindrome or not.
22. Perform following operations on a list of numbers:
 - 1) Insert an element 2) delete an element 3) sort the list 4) delete entire list
23. Display word after Sorting in alphabetical order.
24. Perform sequential search on a list of given numbers.
25. Perform sequential search on ordered list of given numbers.
26. Maintain practical note book as per their serial numbers in library using Python dictionary.
27. Perform following operations on dictionary
 - 1) Insert 2) delete 3) change
28. Check whether a number is in a given range using functions.

29. Write a Python function that accepts a string and calculates number of upper case letters and lower case letters available in that string.
30. To find the Max of three numbers using functions.
31. Multiply all the numbers in a list using functions.
32. Solve the Fibonacci sequence using recursion.
33. Get the factorial of a non-negative integer using recursion.
34. Write a program to create a module of factorial in Python.

Course Learning Outcomes: After studying this course students will be able to:

1. Explain environment, data types, operators used in Python.
2. Compare Python with other programming languages.
3. Outline the use of control structures and numerous native data types with their methods.
4. Design user defined functions, modules, files, and packages and exception handling methods.
5. Learn to handle exceptions in Python.

Text / Reference Books:

AUTHOR	TITLE	Publisher	Year of publication	ISBN
Programming in Python	Programming in Python	BPB	2017	978-9386551276
R. Nageswara Rao	Core Python Programming	Dreamtech Press	2021	978-9390457151
Martin C. Brown	Python, The complete Reference	Tata Mc. Graw Hill	2018	978-9387572942
A. Martelli, A. Ravenscroft, S. Holden	Python in a Nutshell	Shroff/O'Reilly	2017	978-9352135400

Programme structure for B.Sc. (H) Microbiology- 3 years (6th Semester)

Sr. No	Course Code	Course Title	Course Type	Credits					Credit Units
				L	T	P	FW	SW	
1	BCH302	Regulation of Gene Expression	Core Course	4	0	2	0	0	6
2		Synthetic Biology	Core Course	4	0	2	0	0	6
3.		<u>Students will choose any two of the given choices*</u>	Specialization Elective Course	4	0	0	0	0	4
4.	MBO304	1. Clinical Microbiology	Specialization Elective Course	4	0	0	0	0	4
	BIF301	2. Introductory Bioinformatics							
	MBO305	3. Industrial Microbiology							
		4. Microbial Quality Control in Food and Pharmaceutical Industries							
		5. MOOC							
5.		<u>Students will choose any two of the given choices**</u>	Skill Enhancement Course						
		1. Biosensors		2	0	0	0	0	2
		2. Biomaterials		2	0	0	0	0	2
	CBA301	3. BioEntrepreneurship							
		4. MOOC							

Total Credits

24

*The Specialization Elective Courses of 5th and 6th semesters will be pooled together.

**The Skill Enhancement Courses of 5th and 6th semesters will be pooled together.

The Specialization Elective Course and the Skill Enhancement Course can also be taken through MOOC. A maximum of 4 credits per semester can be taken through MOOC.

BCH302: Regulation of Gene Expression

L	T	P	Total Credits
4	0	2	6

Course Objectives: An understanding of different ways prokaryotes and eukaryotes regulate the expression of various gene and methods to study DNA-Protein interaction

Course content and syllabus

	Teaching Hours
Unit I: Regulation of Gene Expression in Prokaryotes	18 hrs
<p>Regulation of Gene Expression in Prokaryotes: concept of operon, ORF. Control at initiation of transcription. Promoter strength and role of sigma factors. Lac Operon (Genetic and Biochemical aspects), araBAD operon. Catabolite repression. trp and his Operons.</p> <p>Regulation of genes for ribosomal RNA and proteins. Bacterial viruses (Lytic and Lysogenic modes).</p> <p>Role of small molecules and RNA in gene control. Riboswitches and bacterial two component system.</p>	
Unit II: Regulation of Gene Expression in Eukaryotes	18 hrs
<p>Regulation of Gene Expression in Eukaryotes: Gene regulation in Yeast (Galactose metabolism, Gal 4 protein, Mating Type), role of mediators, enhancer elements.</p> <p>Chromatin remodelling: histone modification, epigenetic changes</p> <p>Post-transcriptional regulation. RNA silencing: siRNA, miRNA, transitive RNAi, ncRNA. Regulation at translational level</p>	
Unit III: DNA-Protein Interaction	18 hrs
<p>Structures of DNA binding domain: HTH, wHTH, zinc fingers, leucine zippers, HL Loop-sheet-helix. Specificity in DNA-protein interactions.</p> <p>Techniques to study DNA-protein interaction- DNA footprinting, DNA pull down, EMSA, Super-shift, CHIP, reporter assays, Co-crystal studies, yeast two hybrid system, FISH.</p>	
Unit IV:	18 hrs

Genomic regulatory domains: Introduction to regulation of expression of gene clusters locus control region (LCR): structure and function LCR of mouse globin gene cluster Insulators, structure and functions, the insulators of <i>hsp70</i> genes of <i>Drosophila melanogaster</i> , Genomic imprinting of <i>Igf-2</i> and <i>H-19</i> genes	
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List of Experiments -with basic instructions (Total Teaching = 72 hrs)

1. Extraction of total nucleic acids from plant tissue.
2. Diauxic growth curve effect.
3. Isolation of mRNA from yeast by affinity chromatography.
4. Effect of inhibitors on protein synthesis.
5. Accumulation of protein due to proteasome inhibitors.

Course Learning Outcomes:

1. Will have the knowledge of structure and function of genes
2. Concept and knowledge of different strategies in regulation of gene expression in prokaryotes and eukaryotes
3. Understand structure of DNA-binding domains
4. Learn various techniques to study DNA-protein interaction

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Krebs, J.E., Goldstein, E.S., and Kilpatrick, S.T.	Lewin's Genes XII	Jones and Bartlett Learning	12 th Ed		
Watson JD, Baker TA, Bell SP, Gann A, Levine M, Losick R.	Molecular Biology of the Gene	Pearsons Publishers	7 th Ed.	978-9332585478	912

Tropp, B.E.	Molecular Biology Genes to proteins	Jones and Bartelet 4 th Ed	4 th Ed	978-93-80853- 49-9	1096
Alberts, B., Johnson, A., Lewis, J., Morgan, D., Raff, M., Roberts, K., and Walter, P.	Molecular Biology of The Cell	Garland Science	6 th Ed	978-0-8153- 4464-3	1342

Synthetic Biology

L	T	P	Total Credits
4	0	2	6

Course Objectives: Explore synthetic biology to understand the principles of designing and engineering biological systems, enabling the development of novel biotechnological solutions and advancing knowledge in genetic engineering

Course content and syllabus

	Teaching Hours
Unit I: Fundamentals of Synthetic Biology	18 hrs
Modern techniques of DNA assembly – NEBuilder HiFi DNA Assembly, Gibson Assembly, BioBrick Assembly, Golden Gate Assembly. Synthetic bacterial chromosome, synthetic yeast chromosomes for modular metabolic engineering, Genomic engineering using transposable elements in vertebrates	
Unit II: Synthetic Networks	18 hrs
Biological parts – Sensor Proteins (switches), Regulatable promoters, Models of gene expression, artificial networks, production of simple networks capable of producing geneticoscillators and toggle switches, consequences of gene expression variability, examples of synthetic networks – Biofuels and green chemicals.	
Unit III: Fundamentals of System Biology	18 hrs
Stochastic gene expression in prokaryotes and eukaryotes- extrinsic and intrinsic noise, re-wiring of genetic networks to perform cellular functions, Identification of functional units ("network motifs") within large gene interaction networks, a classic study of variability in bacterial gene expression, a classic study of variability in cultured cells, and quantitative PCR-based methods to count mRNAs in individual cells	
Unit IV: Synthetic Proteins	18 hrs

Expanding the chemistry of life by cell free protein synthesis and incorporation of nonnatural amino acids, Engineering of membrane proteins that responds to physical stimuli and their applications: Light-gated channels and pumps for optogenetics, Mechanoreceptors, Temperature- and magnetic field-gated channels. Genetically encoded nanosensors, Ratiometric and intensimetric nanosensors. *In vivo* use of nanosensors

List of Experiments - with basic instructions (Total Teaching = 72 hrs)

1. Cloning of a complete gene expression system by HIFI DNA assembly.
2. Online data mining to identify genes which responds to physical and chemical stimuli.
3. Online data mining to identify promoters which responds to physical and chemical stimuli.
4. in-silico design of a protein with new features.

Course Learning Outcomes:

1. Understand basic concepts of synthetic biology.
2. Learn to construct artificial gene networks and proteins.
3. Learn the techniques to re-wire genetic networks.
4. To know the areas of applications of synthetic biology.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Edited By: Paul S Freemont, and Richard I Kitney	Synthetic Biology — A Primer	World scientific	2012	978-1-84816- 863-3	196
Uri Alon,	An Introduction to Systems Biology: Design Principles of Biological Circuits	Chapman & Hall/CRC	2006	139781588 86426	320

MBO304: Clinical Microbiology

L	T	P	Total Credits
4	0	0	0

Course content and syllabus

Course Objectives: This course is a basic introduction to principle of disease and epidemiology, etiology of infectious diseases caused due to microbial pathogens; and knowledge about the available diagnostic methods and their principles.

	Teaching Hours
Unit I: Introduction about disease Epidemiology and host pathogen interaction	18 hrs
<p>Normal microflora of the human body: Basic concepts of normal microflora of skin, throat, gastrointestinal tract, urogenital tract</p> <p>Host pathogen interaction: Basic Concepts about Infection, Invasion, Virulence, Pathogen, Pathogenicity, Opportunistic infections, Nosocomial infections. Transmission of infection, Pathophysiologic effects of LPS.</p> <p>Sample collection, transport and diagnosis: Method of transportation of clinical samples, principles of different diagnostic tests (ELISA, Immunofluorescence, Agglutination based tests, Complement fixation, PCR, DNA probes).</p>	
Unit II: Bacterial and Viral diseases	18 hrs
<p>Bacterial diseases: Basic Introduction of the following diseases with Symptoms, causes and treatment.</p> <p>Respiratory Diseases: <i>Streptococcus pyogenes, Mycobacterium tuberculosis</i></p> <p>Gastrointestinal Diseases: <i>Escherichia coli, Vibrio cholerae, Helicobacter pylori</i></p> <p>Others: <i>Staphylococcus aureus, Bacillus anthracis, Clostridium tetani, Clostridium difficile</i></p> <p>Viral diseases: Introduction about the following viral disease with Symptoms, mode of transmission and control: Polio, Hepatitis, Rabies, Dengue, AIDS, Influenza, Chikungunya,</p>	
Unit III: Protozoan diseases and Fungal diseases	18 hrs
<p>Protozoan diseases: Introduction about the following protozoan diseases with Symptoms, mode of transmission and control: Malaria, Kala-azar.</p>	

<p>Fungal diseases: Brief description of each of the following types of mycoses and one representative disease to be studied with respect to transmission, symptoms and prevention.</p> <p>Cutaneous mycoses: Tinea pedis (Athlete's foot).</p> <p>Systemic mycoses: Histoplasmosis. Opportunistic mycoses: Candidiasis</p>	
Unit IV: Antimicrobial agents: General characteristics and mode of action	18 hrs
<p>Antibacterial agents: Basic Introduction about modes of action with one example each: Inhibitor of nucleic acid synthesis; Inhibitor of cell wall synthesis; Inhibitor of cell membrane function; Inhibitor of protein synthesis; Inhibitor of metabolism.</p> <p>Antifungal agents: Mechanism of action of Amphotericin B,</p> <p>Antiviral agents: Mechanism of action of Amantadine, Acyclovir, Azidothymidine</p> <p>Antibiotic resistance, MDR, XDR, MRSA, NDM-1.</p>	

Course Learning Outcomes: at the end of the course the students will:

1. Understand the principle of disease and epidemiology, etiology of infectious diseases.
2. Perceive basic knowledge about various bacterial and viral diseases
3. Perceive knowledge about various protozoan and fungal diseases
4. Understand the general characteristics and mechanism of action of different antimicrobial agents.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Ananthanarayan R. and Paniker C.K.J.	Textbook of Microbiology	University Press Publication	11 th ED/2009	9389211433, 978- 9389211436	680
Brooks G.F., Carroll K.C., Butel J.S., Morse S.A. and Mietzner, T.A.	Jawetz, Melnick and Adelberg's Medical Microbiology	McGraw Hill Publication	26 th ED/ 2013	007181292X, 97800718129 2 4	864

Willey JM, Sherwood LM, and Woolverton CJ.	Prescott, Harley and Klein's Microbiology. 9 th Edition	McGraw Hill Higher Education	2013	0073402400, 97800734024 0 6	1152
Madigan MT, Martinko JM, Dunlap PV and Clark DP.	Brock Biology of Microorganism ms. 14 th edition.	Pearson International Edition	14 th ED/ 2014	0321897390, 978- 0321897398	1032

BIF301: Introductory Bioinformatics

L	T	P	Total Credits
4	0	0	0

Course content and syllabus

Course Objectives: Equipping students with foundational knowledge in bioinformatics, enabling them to analyze biological data, navigate bioinformatics tools and databases, and understand the interdisciplinary aspects of this field, thereby preparing them for further studies or careers in bioinformatics and related areas

	Teaching Hours
Unit I: Introduction to Bioinformatics and Biological Databases	18 hrs
Introduction to Bioinformatics. Historical background. Scope of bioinformatics in modern research	
Introduction to biological databases - primary, secondary and composite databases, NCBI, PubMed, nucleic acid databases (GenBank, EMBL, DDBJ, NDB), protein databases (UniProt-Swiss-Prot, PDB), Structure visualization softwares (RasMol, PDBviewer), file formats (FASTA, ASN Genbank).	
Unit II: Sequence alignment	18 hrs
Concepts of sequence similarity, identity and homology. Alignment – local and global alignment, pairwise and multiple sequence alignments, amino acid substitution matrices(PAM and BLOSUM). Programs for pairwise and multiple sequence alignment (CLUSTALW), Introduction to database searching BLAST.	
Unit III: Protein Structure Prediction	18 hrs
Hierarchy of protein structure - primary, secondary and tertiary structures Structural Classes, Motifs, Folds and Domains.	
Protein secondary structure prediction	
Protein tertiary structure prediction in presence and absence of structure template. Energy minimizations and evaluation by Ramachandran plot.	
Protein structure and rational drug design.	
Unit IV: Genome Organization and analysis	18 hrs

Diversity of Genomes: Viral, prokaryotic & eukaryotic genomes. Genome, transcriptome, proteome, 2-D gel electrophoresis, MALDI- TOF spectrometry. Major features of completed genomes: <i>E.coli</i> , <i>S.cerevisiae</i> , <i>Arabidopsis</i> , Human.	
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Course Learning Outcomes:

1. Understand role of biological databases and download appropriate literature, sequences and other relevant information from biological databases
2. Understand importance of sequence alignment
3. Predict structures of proteins
4. Understand organization of genomes and techniques used to study.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Xiong, J.	Essential Bioinformatics	Cambridge University Press	2006	0521706106, 978-0521600828	352
Ghosh, Z. and Mallick, B.	Bioinformatics –Principles and Applications	Oxford University Press	2008	0195692306, 9780195692303	560

MBO305: Industrial Microbiology

L	T	P	Total Credits
4	0	0	4

Course content and syllabus

Course Objectives: This course teaches students about utilization of various microbes in industries, principles of fermentation technology and application of microbes in various industries

	Teaching Hours
Unit I: Introduction to industrial microbes	18 hrs
Microorganisms of industrial importance – yeasts (<i>Saccharomyces cerevisiae</i>), moulds (<i>Aspergillus niger</i>) bacteria (<i>E.coli</i>), actinomycetes (<i>Streptomyces griseus</i>). Industrially important Primary and secondary microbial metabolites. Screening techniques. Techniques involved in selection of industrially important metabolites from microbes.	
Unit II: Industrial fermentation methods	18 hrs
Theory and principles of industrial fermentation, Batch, fed-batch and continuous cultures, Microbial growth and product formation kinetics, media formulation and sterilization, isolation, preservation and improvement of industrially important microorganisms, inoculum development for industrial fermentations, fermenter design, various types of fermenters used in industrial fermentation. Surface, submerged and solid-state fermentation processes. Basic principal of microbial fuel cells and its application .	
Unit III: Industrial applications	18 hrs
Microorganisms involved in Pharma and therapeutic enzymes. Enzymes used in detergents, textiles and leather industries. Production of amylases and Proteases. Production of therapeutic enzymes. Role of microorganisms in bioleaching and textile industry.	
Unit IV: case studies	18 hrs
Industrial microorganisms: cell growth, microbial growth kinetics, factors affecting growth, basic nutrition, principles of production media, components of media, chemical composition of media. Microbial production of Industrial products: Citric acid, Ethanol, Penicillin, Glutamic acid, and vitamin B12.	

Course Learning Outcomes:

1. Understanding fermentation techniques, bioreactor design and media optimization
2. Upstream process for organic acids, alcohols, wine and vinegar
3. Downstream process (Recovery and purification) of antibiotics, enzymes, amino acids and steroids
4. Significance of microbially derived bioproducts for benefit of mankind

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Pelczar, M.J. Jr., Chan ECSand Krieg, N.R.	Microbiology: Concepts and Applications	New York; Madrid : McGraw- Hill,	1993	0070492581, 978007049258 5	957
Cappucino, J.G.	Microbiology- A laboratory manual, 4th ed., Harlow, Addition- Wesley.	Hoboken, N.J.: Pearson	2020	0135188997, 978013520399 6, 01352039	541

Microbial Quality Control in Food and Pharmaceutical Industries

L	T	P	Total Credits
4	0	0	4

Course content and syllabus

Course Objectives: Providing essential knowledge in maintaining microbial quality control within food and pharmaceutical sectors, ensuring adherence to industry standards and regulations, and equipping students for effective application in their professional roles.

	Teaching Hours
Unit I: Microbiological Laboratory and Safe Practices	18 hrs
Good laboratory practices - Good laboratory practices, Good microbiological practices Biosafety cabinets – Working of biosafety cabinets, using protective clothing, specification for BSL-1, BSL-2, BSL-3. Discarding biohazardous waste – Methodology of Disinfection, Autoclaving & Incineration	
Unit II: Determining Microbes in Food / Pharmaceutical Samples	18 hrs
Culture and microscopic methods - Standard plate count, Most probable numbers, Direct microscopic counts, Biochemical and immunological methods: Limulus lysate test for endotoxin, gel diffusion, sterility testing for pharmaceutical products. Molecular methods - Nucleic acid probes, PCR based detection, biosensors.	
Unit III: Pathogenic Microorganisms of Importance in Food & Water	18 hrs
Enrichment culture technique, Detection of specific microorganisms - on XLD agar, Salmonella Shigella Agar, Mannitol salt agar, EMB agar, McConkey Agar, Saboraud Agar Ascertain microbial quality of milk by MBRT, Rapid detection methods of microbiological quality of milk at milk collection centres (COB, 10 min Resazurin assay)	
Unit IV: HACCP for Food Safety and Microbial Standards	18 hrs
Hazard analysis of critical control point (HACCP) - Principles, flow diagrams, limitations Microbial Standards for Different Foods and Water – BIS standards for common foods and drinking water	

Course Learning Outcomes:

1. Develop skills on disinfection of instruments and equipments in laboratory and Hospitals.
2. Develop knowledge and skills on microbiological laboratory safety- General rules and regulations

3. To impart knowledge on different culture techniques.
4. To learn Microbial Standards for Different Foods and Water.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Pelczar, M.J. Jr., Chan ECSand Krieg, N.R.	Microbiology: Concepts and Applications	New York; Madrid : McGraw- Hill,	1993	0070492581, 978007049258 5	957
Cappucino, J.G.	Microbiology- A laboratory manual, 4th ed., Harlow, Addition- Wesley.	Hoboken, N.J.:Pearson	2020	0135188997, 978013520399 6, 0135203996	541

Biosensors

L	T	P	Total Credits
2	0	0	2

Course content and syllabus

Course Objective: Offering fundamental understanding in the design, development, and application of biosensors, and preparing students to employ these advanced technologies across diverse fields such as healthcare, environmental monitoring, and biotechnology.

	Teaching Hours
Unit I: Introduction to Biosensors	9 hrs
Introduction to biosensors Biosensor classification Main elements in biosensors. Properties of Biosensors-sensitivity, selectivity and stability.	
Unit II: Biomolecules in biosensors:	9 hrs
Biomolecules in biosensors: DNA, enzyme, antibody, antigen, protein, peptide, aptamer, Protein/antibody-based sensors: protein immobilisation, specificity, binding constants, kinetics, diffusion.	
Unit-III: Basics of detection methods	9 hrs
Basics of detection methods: Fluorescence Spectroscopy, UV-Vis Absorption and Emission, Surface Plasmon Resonance, Magnetic labeling, Electrochemical Detection. Fundamentals of transducers.	
Unit IV: Applications of biosensing technology	9 hrs
Point-of-care diagnosis, molecular diagnostics, Fluorescence and colorimetric biosensors, microfluidics and paper-based diagnostics Detection of viruses and bacteria, Microbial biosensor. Applications of nanotechnology in biosensors.	

Course Learning Outcomes:

1. Perceive knowledge about the biosensor and its classification Understand the working principle of biosensor.
2. Perceive knowledge about different elements of biosensor and their applications in diagnostics.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
Donald G. Buerk	Biosensors: Theory and Applications	CRC Press,	2009	ASIN : B07CSVY2HM	232
Brian R. Eggins	Chemical Sensors and Biosensors	John Wiley & Sons	2002	978-0-471-89914-3	300
Ashutosh Tiwari, Anthony P.F. Turner	Biosensors Nanotechnology	Scrivener Publishing LLC	2014	9781118773512	533

Biomaterials

L	T	P	Total Credits
2	0	0	2

Course Objective: To impart knowledge on structure-property relationship in biomaterials and their applications as implants.

Course content and syllabus

	Teaching Hours
Unit I: Introduction	9 hrs
Materials-Bulk properties and surface properties	
Unit II: Material Classes	9 hrs
Class of materials used in biomedical applications	
Unit III: Cell-Material Interactions	9 hrs
Biological interactions with materials-Proteins, cells, and tissues, biological responses: Inflammation, immunity, toxicity, coagulation, tumorigenesis. Biofilms, Pathological calcification, Biocompatibility	
Unit IV: Applications	9 hrs
Applications of biomaterials: drug delivery, tissue engineering, cardiovascular, orthopedic, dental, functional tissues, etc.	

Course Learning Outcomes:

1. Students will be able to understand the fundamentals and classes of materials.
2. Describe interactions between biomaterials, proteins and cells.
3. Explain methods to modify surfaces of biomaterials and choose material for desired biological response.
4. Analyse the interactions between biomaterial and tissue for short term and long-term implantations, distinguish between reactions in blood and in tissue.

Text / Reference Books:

Author	Title	Publisher	Ed/year	ISBN No	Pages
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Buddy D. Ratner, Allan S. Hoffman, Frederick J. Schoen, Jack E. Lemons	Biomaterials Science: An Introduction to Materials in Medicine	Academic Press, USA	2004	0123746264, 978-0123746269	1573
J.B. Park and J.D. Bronzino	Biomaterials: Principles and Applications	CRC Press	2002	0849314917, 9780849314919	250
K.C. Dee, D.A. Puleo and R. Bizios	An Introduction to Tissue-Biomaterial Interactions	Wiley	2002	0471253944, 9781280366642	228
T. M. Wright, and S. B. Goodman	Implant Wear in Total Joint Replacement: Clinical and Biologic Issues, Material and Design Considerations	American Academy of Orthopaedic Surgeons	1 st ED/ 2001	0892032618, 9780892032617	224
Park J.B. and Lakes R.S	Biomaterials: An Introduction, 3 rd edition	Springer press	3 rd ED/ 2007	978-0387378794	574

CBA301: BioEntrepreneurship

L	T	P	Total Credits
2	0	0	2

Course Objectives: To help students gain understanding of the basic concepts of entrepreneurship, diagnose new business opportunities, formulate business plans, and identify different institutional support available to the entrepreneurs.

Course Content/ Syllabus

	Teaching Hours
Unit I: Basic Concepts of Entrepreneurship	9
Introduction to Entrepreneurship: Meaning, Background, Importance, The Benefits of Entrepreneurship, The Potential Drawbacks of Entrepreneurship, Factors that Influence Entrepreneurship, How to Avoid the Pitfalls, Factors Responsible for Entrepreneurship Growth; Entrepreneur Background and Characteristics; Entrepreneurial Potential in a Prospective Entrepreneur; Entrepreneurial Skills and Competencies; Types of entrepreneurs and entrepreneurship, Myths and Realities about Entrepreneurs; New Trends in Entrepreneurship Development; Economic Development through Entrepreneurship; Role of Entrepreneurship in the Economic Development of India	
Unit II: Environmental Monitoring and Importance of Business Idea	9
Creativity and innovation, Role of Creativity & Innovation in Entrepreneurship, Sources of New Ideas – Consumers, Existing Products and Services, Distribution Channels, Federal Government, Research and Development; Methods of Generating Ideas – Focus Groups, Brainstorming, Brainwriting, Problem Inventory Analysis; Creative Problem Solving – Brainstorming, Reverse Brainstorming, Brainwriting, Gordon Method, Checklist Method, Free Association, Forced Relationships, Collective Notebook Method, Attribute Listing Method, Big-dream Approach, Parameter Analysis, Mind Mapping, Force-Field Analysis, TRIZ, Rapid Prototyping; Innovation, Types of Innovation – Breakthrough, Technological, and Ordinary Innovation; Opportunity Recognition; Product Planning and Development Process – Idea Stage, Concept Stage, Product Development Stage, Test Marketing Stage, and Commercialization Stage; Technology Readiness Levels; Intellectual Property Rights	

Unit III: Scanning the Environment & Business Plan Development	9
Identifying the business opportunity: SWOT and PESTEL analysis, Viability Screening/Feasibility Analysis: Market Feasibility, Production Feasibility, Organisational Feasibility, Financial Feasibility; Business Plan Development: Introduction, Business Plan, Various Business Models – The Business Model Canvas, The Lean Canvas, Types of Business Plans, Structure of a Basic Business Plan, Creating a Business Plan – Executive Summary, General Company Description, The Opportunity or Competitive Analyses, Market Research and Industry Analysis, Strategy, The Team, Marketing Plan, Operational Plan, Financial Plan, and An Appendix	
Unit IV: Sources of Capital and Institutional Support for Entrepreneurs	9
Sources of Funding for Entrepreneurs: Bootstrapping, Friends and Family Members, Crowdfunding, Angel Investment, Venture Capital, Financial Institutions, Bank Loans, Trade Credit, Initial Public Offerings/Issue of Shares, Debentures; Role of Government in Promoting Entrepreneurship: Atal Innovation Mission, Biotechnology Industry Research Assistance Council, Department of Science and Technology, Digital India, Jan Dhan-Aadhaar-Mobile, Make in India, National Skill Development Mission, Pradhan Mantri Kaushal Vikas Yojana, Science for Equity Empowerment and Development, Stand-Up India, Start-Up India, Support to Training and Employment Programme for women, Trade-Related Entrepreneurship Assistance and Development, USAID	

Course Learning Outcomes: On completion of the course, the student shall be able to:

1. Understand the concept of entrepreneurship, its emergence and its need for society.
2. Formulate a business idea and diagnose for a new business opportunity.
3. Identify various business gaps and develop a business plan
4. Evaluate and identify different institutional support available to the entrepreneur.

List of Professional Skill Development Activities (PSDA):

1. Research on growth profile of an entrepreneur
2. Identify opportunity, generate idea and conduct feasibility Analysis
3. Design a Business Plan
4. Develop an Entrepreneur Journal where reflection and personal experiences will be recorded
5. Write personal insights, lessons learned, other readings, and the video clips you watch in this semester
6. Interview one entrepreneur mentor and come up with five good business questions you would like to ask him or her

7. Comparative study of startups in the field of Biopharmaceuticals, Bioagriculture, Bioindustry, and Bioservices.

Text / Reference Books:

Author	Title	Publisher	Year of publication	ISBN	Pages
Evan J. Douglas	Entrepreneurial Intention: Past, Present, and Future Research	Edward Elgar Publishing	2020	978-1-78897-522-3	216
Justin G. Longenecker, J. William Petty, Leslie E. Palich, and Frank Hoy	Small Business Management: Launching & Growing Entrepreneurial Ventures (20 th Edition)	Cengage	2023	978-0-3577-1880-3	712
Mike Kennard	Innovation and Entrepreneurship	Routledge	2021	978-0-367-51057-2	114
Debashish Biswas and Chanchal Dey	Entrepreneurship Development in India	Routledge	2021	978-0-367-76219-3	117
Robert D. Hisrich, Micheal P. Peters, Dean A. Shepherd, Sabyasachi Sinha	Entrepreneurship (11 th Edition)	McGraw Hill	2020	978-9390113309	696
Donald F. Kuratko and Jeffrey S. Hornsby	New Venture Management: The Entrepreneur's Roadmap for Development, Management, and Growth (3 rd Edition)	Routledge	2020	978-0367466725	356

Bruce R. Barringer and R. Duane Ireland	Entrepreneurship: Successfully Launching New Ventures (6 th Edition)	Pearson	2019	978-1-292-25533-0	617
Norman M. Scarborough and Jeffrey R. Cornwall	Essentials of Entrepreneurship and Small Business Management (9 th Edition)	Pearson	2019	978-1-292-26602-2	827
Mary Jane Byrd and Leon Megginson	Small Business Management: An Entrepreneur's Guidebook (8 th Edition)	McGraw Hill	2017	978-1259538988	496
Robert D. Hisrich and Veland Ramadani	Effective Entrepreneurial Management: Strategy, Planning, Risk Management, and Organization	Springer	2017	978-3-319-50465-0	230
Stephen Spinelli, Jr. and Robert J. Adams, Jr.	New Venture Creation: Entrepreneurship for the 21st Century (10 th Edition)	McGraw-Hill Education	2016	978-0-07-786248-8	484
David H. Holt	Entrepreneurship: New Venture Creation	Pearson	2016	978-9332568730	584
Peter F. Drucker	Innovation and Entrepreneurship	Harper Business	2006	978-0060851132	288

Robert J. Calvin	Entrepreneurial Management	McGraw-Hill	2005	9780071450928	295
Steve Mariotti	Entrepreneurship and Small Business Management	Pearson publishers	2014	978-0133767186	